

# Quantitative Disease Resistance to White Pine Blister Rust at Southwestern White Pine's (Pinus strobiformis) Northern Range

Jeremy S. Johnson<sup>1, 2, 3\*</sup>, Richard A. Sniezko<sup>2</sup>

<sup>1</sup>Department of Environmental Studies, Prescott College, United States, <sup>2</sup>USDA Forest Service, Dorena Genetic Resource Center (DGRC), United States, <sup>3</sup>School of Forestry, Northern Arizona University, United States

Submitted to Journal: Frontiers in Forests and Global Change

Specialty Section: Pests, Pathogens and Invasions

Article type: Original Research Article

*Manuscript ID:* 765871

Received on: 27 Aug 2021

Revised on: 30 Sep 2021

Journal website link: www.frontiersin.org



#### Conflict of interest statement

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest

#### Author contribution statement

JSJ analyzed and interpreted the data, and led the writing of the manuscript, RAS conceived of the paper assisted with writing, analysis, and interpretation. Both authors reviewed and edited the manuscript and approved of the final version.

#### Keywords

Cronartium ribicola, Pinus strobiformis, bark reactions, five needle pines, quantitative disease resistance, white pine blister rust

#### Abstract

#### Word count: 300

White pine blister rust, caused by the non-native, invasive fungal pathogen Cronartium ribicola, is a significant cause of mortality in white pines (Pinus subgenus Strobus) in North America. Along with climate-driven range contraction, mortality from blister rust can seriously impact the abundance and distribution of the nine white pine species native to the United States and Canada. Very little evaluation of this disease in southwestern white pine (Pinus strobifomis) has been previously undertaken, but genetic resistance to the disease has been documented, including major gene resistance (MGR) conferred by a dominant R gene. Data is emerging suggesting that the species also has quantitative disease resistance (QR). Our results suggest QR occurs at low frequency, with perhaps 10% of trees having a moderate level (>35% survival). We assess progeny arrays from 40 P. strobiformis families (1873 seedlings), originating from three populations, inoculated with C. ribicola. Subsequently, the seedlings were assessed for signs, symptoms and resulting impact in a common garden trial over a 7.5-year period to determine the types and frequency of resistance in a portion of this species' range. There was a high incidence of both stem symptoms and mortality in the P. strobiformis families tested, and families ranged in survival from 0 to 84.6%. Three families had >70% survival, representing perhaps the highest documented OR to date in a North American white pine species. Approximately 7% of surviving seedlings showed no stem symptoms, with approximately 16% of seedlings surviving with infections of generally low severity. QR traits associated with improved survival were primarily related to lower severity of infection, a reduced number of stem symptoms, and an increased number of bark reactions. Despite the high overall susceptibility, the presence of QR appears to be at a frequency and level useful to forest managers involved in restoration and reforestation efforts.

#### Contribution to the field

White pine blister rust is a devastating non-native invasive disease that impacts nearly all North Americas white pines. Our study tracks and quantifies the impact that multiple resistance phenotypes have on the overall survival of infected seedlings of one of these pines (Pinus strobiformis) and provides a glimpse at the range of quantitative disease resistance that may be present in this species. Our results are one of the first real breakdowns of quantitative disease resistance in any species of white pine and provide encouragement and cautious optimism for foresters and land managers as they plan for the ever-expanding advancement of the disease white pine blister rust.

#### Funding statement

JSJ was supported by NSF 1340852. Dave Conklin, USFS R3 FHP coordinated the cone collections with others in Region 3, and Region 3 FHP provided funding for the study.

#### Ethics statements

#### Studies involving animal subjects

Generated Statement: No animal studies are presented in this manuscript.

#### Studies involving human subjects

Generated Statement: No human studies are presented in this manuscript.

#### Inclusion of identifiable human data

Generated Statement: No potentially identifiable human images or data is presented in this study.



#### Data availability statement

Generated Statement: The original contributions presented in the study are included in the article/supplementary material, further inquiries can be directed to the corresponding author/s.



# Quantitative Disease Resistance to White Pine Blister Rust at Southwestern White Pine's (*Pinus strobiformis*) Northern Range

1 Jeremy S. Johnson<sup>1,2,3\*</sup>, Richard A. Sniezko<sup>2</sup>

- <sup>2</sup> <sup>1</sup>Prescott College, Department of Environmental Studies, Prescott, AZ, USA
- <sup>3</sup> <sup>2</sup> Dorena Genetic Resource Center, USDA Forest Service, Cottage Grove, OR, USA
- 4 <sup>3</sup> Northern Arizona University, School of Forestry, Flagstaff, AZ, USA
- 5 \* Correspondence:
- 6 Corresponding Author
- 7 jeremy.johnson@prescott.edu

8 Keywords: Cronartium ribicola, Pinus strobiformis, bark reactions, five needle pines,

9 quantitative disease resistance, white pine blister rust.

### 10 Abstract

- 11 White pine blister rust, caused by the non-native, invasive fungal pathogen *Cronartium*
- *ribicola,* is a significant cause of mortality in white pines (*Pinus* subgenus *Strobus*) in North
- 13 America. Along with climate-driven range contraction, mortality from blister rust can
- 14 seriously impact the abundance and distribution of the nine white pine species native to the
- 15 United States and Canada. Very little evaluation of this disease in southwestern white pine
- 16 (*Pinus strobifomis*) has been previously undertaken, but genetic resistance to the disease has
- 17 been documented, including major gene resistance (MGR) conferred by a dominant *R* gene.
- 18 Data is emerging suggesting that the species also has quantitative disease resistance (QR).
- 19 Our results suggest QR occurs at low frequency, with perhaps 10% of trees having a
- 20 moderate level (>35% survival). We assessed progeny arrays from 40 P. strobiformis families
- 21 (1873 seedlings), originating from three populations, inoculated with C. ribicola.
- 22 Subsequently, the seedlings were assessed for signs, symptoms and resulting impact in a
- 23 common garden trial over a 7.5-year period to determine the types and frequency of
- 24 resistance in a portion of this species' range. There was a high incidence of both stem
- 25 symptoms and mortality in the *P. strobiformis* families tested, and families ranged in
- 26 survival from 0 to 84.6%. Three families had >70% survival, representing perhaps the
- 27 highest documented QR to date in a North American white pine species. Approximately 7%
- of surviving seedlings showed no stem symptoms, with approximately 16% of seedlings
- 29 surviving with infections of generally low severity. QR traits associated with improved
- 30 survival were primarily related to lower severity of infection, a reduced number of stem
- 31 symptoms, and an increased number of bark reactions. Despite the high overall
- 32 susceptibility, the presence of QR appears to be at a frequency and level useful to forest
- 33 managers involved in restoration and reforestation efforts.

### 34 1 Introduction

35 Non-native invasive pathogens and pests have had a substantial negative impact on tree species and their associated ecosystems. More recently, because of climate change and 36 37 changing patterns of transient disturbances, there are increased concerns about how these 38 pests and pathogens will affect the distribution and stability of forest host species. 39 *Cronartium ribicola* J.C. Fisch, the invasive non-native fungal pathogen responsible for the 40 disease white pine blister rust, has caused high mortality to both economically and 41 ecologically important white pine species since its introduction in North America in the 42 early 20<sup>th</sup> century (Kinloch, 2003). For example, the pathogen, along with early 20<sup>th</sup> century 43 logging and harvest practices, has resulted in extensive forest loss in western white pine 44 (Pinus monticola ex. Don) stands in the Interior West USA where it once comprised 25-50% 45 of the forests in the region (Fins et al., 2002; Tomback and Achuff, 2010). There are eight 46 species of *Pinus* subgenus *Strobus*, known as the white pines or five-needle pines, in the western United States (and three of these also occur in Canada). The white pines broadly 47 48 are keystone species where they are found, are important components of the hydrological 49 cycle, providing erosion control, and wildlife habitat as well as a component of temperate 50 forest biodiversity (Schoettle, 2004; Tomback and Achuff, 2010). All of the North American 51 white pine species are extremely susceptible to white pine blister rust (Hoff et al., 1980; 52 Kinloch, 2003; Sniezko et al., 2008) and all but Pinus longaeva have C. ribicola within their 53 native range in the U.S. and Canada, but the disease has not yet been documented in

54 Mexico or Central America.

55 *Pinus strobiformis* Engelm. (Southwestern white pine) is a large, long-lived conifer native to

56 the southwestern USA and Mexico (Looney and Waring, 2012). The species thrives in

57 mixed-conifer stands at mid to high elevations (Looney and Waring, 2012; Shirk et al., 2018)

58 where it is moderately shade-tolerant (Goodrich and Waring, 2017) and drought-tolerant

59 (Bucholz et al., 2020), often occurring in sky islands. Within the northern extent of its range,

60 *P. strobiformis* is part of a moving hybrid zone with *Pinus flexilis* (Menon et al., 2020) with

61 some evidence of increased fitness due to introgressed alleles from *P. flexilis* (Menon et al.,

62 2021).

63 Like all other North American white pines, *P. strobiformis* is very susceptible to the disease

64 white pine blister rust (Conklin et al., 2009; Sniezko et al., 2011). However, the fungal

65 pathogen is a more recent invader of the southwestern U.S., detected in the Sacramento

66 Mountains of New Mexico in 1990 (Hawksworth, 1990) with a likely earlier arrival in the

67 1970s (Jacobi et al., 2018), and has not yet been identified within the pine's core range in

- 68 Mexico. Unease about the potential impacts of increasing incidence of rust in *P. strobiformis*
- 69 forests, along with projected increasing aridity and range contraction in the southern
- 70 portion of the species range (Seager and Vecchi, 2010; Shirk et al., 2018), have led to an
- 71 increased interest in what type and frequency of genetic resistance may naturally occur in
- the species. Characterizing patterns of genetic resistance will prove valuable, especially in
- the context of identifying how the most affected stands of *P. strobiformis* will be impacted,

- 74 and will inform the conservation, restoration, and management actions that are likely to be
- 75 the most beneficial. To be successful for management and mitigation, genetic resistance
- 76 must be stable, durable and usable at an appropriate frequency and level (Sniezko et al.,77 2020).

78 Despite the ubiquitous susceptibility of white pines to white pine blister rust, and the lack 79 of co-evolution with this non-native disease, white pines generally do have a low frequency 80 of natural genetic resistance (Hoff et al., 1980; King et al., 2010; Sniezko et al., 2014), often 81 referred to as exapted resistance (Gould and Vrba, 1982; Bartholomé et al., 2020). Genetic 82 resistance in white pine species is generally associated with having either a dominant major R gene (MGR), Cr3 in P. strobiformis (Kinloch and Dupper, 2002), conveying complete 83 84 resistance, or quantitative disease resistance (QR), with an array of phenotypic traits and more complex patterns of inheritance (Hoff et al., 1980; Sniezko et al., 2014; Weiss et al., 85 86 2020; Liu et al., 2021). Quantitative disease resistance can prevent some trees from 87 developing cankers, but in other cases the trees do develop cankers, but respond via bark 88 reactions and other mechanisms (Box 1) which reduce the rate of disease development and 89 in some cases halt the spread of the fungus (Hoff, 1986; Sniezko et al., 2014; Vázquez-Lobo 90 et al., 2017). Previous studies have acknowledged several traits potentially associated with 91 QR in white pines including fewer needle spots (Hoff and McDonald, 1980), reduced 92 number of stem symptoms (cankers and bark reactions) (Kegley and Sniezko, 2004; Sniezko 93 et al., 2014), slow fungus growth in the needles (Hoff, 1988), the occurrence of partial or 94 complete bark reactions (Hoff, 1986), slow canker growth (Hunt, 1997), delayed onset of 95 symptom development (Kegley and Sniezko, 2004), and ultimately increased survival 96 (Sniezko et al., 2014; Sniezko et al., 2020). The aforementioned traits are not mutually 97 exclusive, and often occur together. Within and between families there is variation in the 98 expression of different resistance components (Sniezko et al., 2014). 99 Some white pines with MGR exhibit a hypersensitive-like reaction that occurs within the 100 needle and stops the pathogen from spreading into the stem of the tree. While MGR seems 101 to be the best option for stabilizing forests with blister rust hazard, this type of resistance is 102 often overcome by virulent strains of the disease (Kinloch and Comstock, 1981; Kinloch et 103 al., 1999; Kinloch and Dupper, 2002; Kinloch et al., 2004) making it less durable than QR 104 (Sniezko et al., 2020). As such QR represents a more usable form of resistance, especially in

- 105 combination with MGR, in the face of an evolving white pine blister rust pathosystem.
- tos combination with WGK, in the face of an evolving white pine blister fust pathosystem.
- 106 Notwithstanding its promise, QR can be difficult to characterize, in part owed to the time
- 107 commitment needed to monitor trials for several years and the challenge associated with
- 108 interpreting the many interacting resistance phenotypes and the quantitative genetic 109 contributions. Because of these difficulties, the underlying genetic control of OR in wh
- 109 contributions. Because of these difficulties, the underlying genetic control of QR in white110 pines is not well understood (Vázquez-Lobo et al., 2017). There are some emerging
- examples in the literature, mostly centered on *Pinus lambertiana* and *P. monticola* (Kegley
- and Sniezko, 2004; Kolpak et al., 2008; Kinloch et al., 2012; Liu et al., 2013; Sniezko et al.,
- 113 2014; Vázquez-Lobo et al., 2017; Sniezko et al., 2020; Weiss et al., 2020), which have begun to

- 114 characterize QR in white pines, and in some cases the genomic loci that partially contribute
- 115 to this form of resistance (Liu et al., 2020; Weiss et al., 2020; Liu et al., 2021). However, the
- 116 most detailed characterization of QR traits is when progeny arrays (families) grown in a
- 117 common garden are assessed for variation in inheritance of resistance traits and by the co-
- 118 occurrence of one or more of these traits allowing individuals, on average, to survive longer
- 119 (Sniezko, 2006; Sniezko et al., 2014; Sniezko et al., 2020).
- 120 The objective of this study is to assess the phenotypes associated with increased
- 121 survivorship in *P. strobiformis* and begin to characterize QR in the species from several
- 122 populations in the New Mexico portion of the species range. We monitored the
- 123 development of disease symptoms in *P. strobiformis* for 7.5 years post-inoculation. To
- 124 characterize the components of QR to white pine blister rust, we collected measurements
- 125 both within and between families on (1.) disease incidence (the percentage of plants
- 126 infected, both in the needle and the stem, in a seed family) (2.) the number of stem
- symptoms (3.) the percentage of individuals within a family with stem symptoms (4.) the
- 128 percentage of individuals within a family with bark reactions (5.) the timing of stem
- symptom development, (6.) the severity of infection, and (7.) temporal patterns of survival.
- 130 We hypothesize that *P. strobiformis* families with a reduced number of stem symptoms
- 131 (including some seedlings that have none), higher occurrence of either complete or
- 132 incomplete bark reactions, and later appearance of stem symptoms, will lead to higher
- 133 survival, and will indicate the presence of QR.
- 134
- 135 2 MATERIALS AND METHODS

### 136 Experimental Design

137 Cones from 40 open-pollinated, P. strobiformis trees were collected in 2008 from three 138 populations, the Lincoln (n = 20), Santa Fe (n = 10), and Cibola (n = 10) National Forests in 139 New Mexico, USA (Figure 1). Seed from the 40 parent trees were sown in March 2009 at the 140 U.S. Department of Agriculture, Forest Service's Dorena Genetic Resource Center (Cottage 141 Grove, Oregon, USA). Bradford Canyon, on the Lincoln NF, had previously been 142 documented with very high incidence of white pine blister rust, in some stands 143 approximately 90% of the trees have been infected (Conklin, 2004; Conklin et al., 2009). 144 Seventeen parent trees selected from the Lincoln NF, the site where the first P. strobiformis 145 trees with MGR were documented, were canker-free, suggesting putative resistance 146 (designated here as Lincoln R), in addition three trees with moderate-to-heavy cankering 147 were selected to serve as susceptible controls (Lincoln S). The Lincoln R parent trees had 148 been previously screened for MGR at the USDA Institute of Forest Genetics in Placerville, 149 CA and they did not segregate in ratios that would be expected if they carried the *Cr3* allele conveying MGR (D. Conklin Personal Communication). Parent trees selected from the Santa 150 151 Fe NF and Cibola NF were selected from locations with little or no rust present at the time 152 of cone collection and represent random genotype selections (Table 1). Seedlings were 153 grown for two years, one year in 164 cm<sup>3</sup> supercell Cone-tainers<sup>tm</sup> (Ray Leach, Canby,

- 154 Oregon) in family blocks in an unheated greenhouse and then transplanted to 0.9 m x 1.2 m
- 155 x 0.3 m boxes outside for the second growing season. Twelve to 60 seedlings were available
- 156 for each family (mean = 47), and seedlings were transplanted into family row plots in
- 157 randomized complete block design with six blocks and up to 10 seedlings per family per
- block. Seedlings were inoculated in September of 2010 with basidiospores of *C. ribicola*.
- 159 Details of the standard Dorena Genetic Resource Center (GRC) inoculation procedures are
- 160 outlined elsewhere (Kegley and Sniezko, 2004; Sniezko et al., 2008; Sniezko et al., 2011).
- 161 Mean inoculums density was 4,527 spores/cm<sup>2</sup>; basidiospore germination was 98.7%. Both
- 162 primary and secondary needles were present on seedlings at the time of inoculation.
- 163

# 164 Disease trait assessment

165 Following inoculation with *C. ribicola,* infected seedlings were periodically assessed for the 166 presence of rust symptoms, with data collection ending in February 2018 (7.42 years postinoculation) (Table 2). Seedlings that died from causes other than white pine blister rust 167 were removed from analysis (n = 10) and a total of 1883 seedlings were included in the 168 169 study. The first assessment occurred approximately 0.75 years post-inoculation and seven 170 assessments were conducted over the course of the trial. The timing of early assessments is 171 calibrated to capture the peak of trait development (e.g. spots and canker emergence). All 172 seedlings were assessed for a core set of traits. Specifically, number of needle spots at first 173 assessment, the presence/absence of needle spots at second assessment, number of cankers, 174 number of bole infections, number of bark reactions, number of partial bark reactions, 175 overall severity of infection, and survival. Full counts of the number and type of stem 176 symptoms were completed at second assessment and the presence of additional stem 177 symptoms were noted at subsequent inspections, since the growth and merging of cankers 178 made later counts more problematic. The counts represent one point in time, and some 179 seedlings showed stem symptoms at later assessments. Additionally, both pre-inoculation height (recorded after inoculation but before 3rd year growth) (April 2011) and height 180 present one growing season post-inoculation, recorded during the 2<sup>nd</sup> assessment (February 181

182 2012; 1.42 years post-inoculation).

183 Based on the level and severity of infection with white pine blister rust, each tree was also

- 184 assigned a severity classification at each assessment. The classification assigns a seedling a
- numeric value that assesses the severity of damage from 0, no infection, to 9, dead from rust
- 186 with classes designated by the degree to which a canker has encircled the bole of the
- 187 seedling and expanded vertically. For example, a tree that is infected with blister rust
- 188 (presence of needle spots and a canker) with intermediate severity, a normal canker
- encircling >50% but <100% of the bole but little vertical expansion, would receive a rating of
- 190 4. The severity rating and the disease trait phenotypes are standard measurements recorded
- as part of rust inspections at the USDA Dorena GRC (supplemental table S1). The severity
- rating for each seedling is dynamic and can change (increase or decrease) with each
- 193 subsequent assessment, reflecting the degree of rust progression or resistance response.

- 194 Seedlings can have one or many stem symptoms and the severity provides a composite look
- 195 at the progression of all infections present at each point in time.
- 196

### 197 Infection Means

198 Percentages and means were calculated from the measured rust phenotypes at the family, 199 and forest scale. For final tally of needle spots, bole infections, complete bark reactions, 200 partial bark reactions, all bark reactions (BRall), stem symptoms (normal cankers + bark 201 reactions + partial bark reactions), and survival, percentages were calculated by 202 categorically assigning individuals as either symptomatic or not (binary) for a rust 203 phenotype. #Spots, #BI, #BRall, and severity was assessed as the family mean of the 204 individual scored values. Forest means were assessed as the mean of family means for each 205 population. Number of bole infections and normal cankers were assessed at the 2<sup>nd</sup> 206 assessment following inoculation. Existence of stem symptoms was assessed at all following 207 inspections along with mortality, %survival and severity allowing us to assess the temporal progression of the disease and delayed development of traits (survival with rust), a 208 209 potential characteristic of QR. Percent bark reaction was calculated using only individuals 210 within a family that developed stem symptoms to avoid underestimating the trait by 211 including those individuals that remained stem symptom free.

212

213

### Timing and Development of Resistance Traits

214 The temporal pattern, or timing of symptom development can result as a form of QR. If the 215 spread of fungal hyphae through live tissue can be slowed, known as slow fungal growth 216 (SFUG) in the needle (Hoff, 1988) or slow canker growth (SCANK) if in the stem (Hoff and 217 McDonald, 1980; Hunt, 1997), the overall survival can be prolonged. We interpret this delay 218 in symptom development from the percentage of early stem symptoms (%ESS<sub>2\_4</sub>), where the 219 maximum value is 100% when all seedlings in a family with stem symptoms at 2<sup>nd</sup> 220 assessment and the minimum is 0% when no trees have stem symptoms at 2<sup>nd</sup> assessment. 221 The focus is on delayed stem symptom development. %ESS is calculated between 2<sup>nd</sup> (1.42) 222 ypi) and 4th (3.25 ypi) assessment. Families that had a lower %ESS may suggest a form of 223 QR. Additionally, the percentage of seedlings that developed stem symptoms but survived 224 (SSALV) was calculated from the subset of individuals that did develop stem symptoms. 225 Those families with a higher SSALV percentage may also suggest a form of QR.

226 227

### Test of Major Gene Resistance

The presence of stem or branch cankers were aggregated into a cumulative binary measure of the phenotype stem symptom-free (SS-free) or stem symptom (SS) at the final

assessment. Segregation ratios of SS-free:SS were tested to identify the potential presence of

- MGR. We used the Mendelian segregation ratios 1:1 (Rr x rr) and 3:1 (Rr x Rr) for SS-free:SS
- 232 seedlings and tested the hypothesis that each family did not differ significantly from a
- 233 probability of 0.5 or 0.75 respectively using an exact binomial test. Maternal source trees

whose families failed to differ significantly from expected 1:1 or 3:1 ratios are inferred to potentially be heterozygous for the *Cr3* allele and possess MGR.

236 237

# Frequency of Quantitative Disease Resistance

Variation in family performance was used to assess the level of QR. On an individual basis, the most susceptible seedlings were those that were cankered earliest and had the earliest

240 mortality. The most susceptible families (and presumably most susceptible parent trees)

were those in which 100% of seedlings were cankered by 2<sup>nd</sup> assessment and 100% mortality

- 242 by 3<sup>rd</sup> assessment. The most resistant families were those with highest survival (lowest rust
- related mortality) at the final assessment. No one trait fully encompasses the full gamut of
- resistance between the families, but perhaps the most important is survival which
- conceivably could vary from 0 to 100 percent for QR families. Survival potentially includesboth canker-free seedlings as well as seedlings with stem symptoms that are alive (SSALV)
- 247 at later assessments (severity of stem symptoms on these living seedlings also may vary).
- 248 Within the group of highest susceptible families described above, the most susceptible
- 249 might be those with the highest number of Spots1 and/or number of stem symptoms early.
- 250 Classification of QR in *P. strobiformis* families was based on seedling families that survived
- 251 for the duration of the trial even in the presence of rust infection. The percentage of
- surviving individuals within a family at the end of the trial was used as a proxy for the
- 253 frequency of QR that may be found in the field under high rust conditions. Survival was
- 254 further partitioned into all survivors (RSurv), survivors with infection (SSALV), and
- survivors that were stem symptom free (SS-free). An additional point is the timing and
- 256 progression of stem symptoms and mortality. By contrast, MGR is typically defined using a
- 257 single trait, cankered vs non-cankered.
- 258 Differences in mean disease symptom between both families and National Forests was
- assessed using analysis of variance (ANOVA) at 95% confidence. Regression analysis on
- traits was carried out using linear mixed effects models fit by REML with random factors of
- 261 parent trees nested within Forest Stand. All statistical analyses were carried out in the R
- 262 statistical environment (RCoreTeam, 2020).
- 263

# 264 **3 RESULTS**

265

# 266 Characterizing QR Resistance Traits

267 Overall susceptibility to white pine blister rust in progenies from native stands was high.

268 Inoculation was successful and indicated by nearly all seedlings developing needle spots on

269 secondary needles (99.64% at 0.75 years post-inoculation). Family variation in needle spots

270 ranged between 95.83% to 100%, and the number of spots varied dramatically by family

- 271 (overall mean number of needle spots per family was 41.00 with family means ranging from
- 14.19 to 84.05 (table 3). By 1.42 years-inoculation (2<sup>nd</sup> assessment) the percent needle spots

- 273 per family was somewhat lower, at 80.03% with a family range between 50.0 to 100%
- 274 (Supplemental Data Table S2), suggesting possible resistance related needle shed (*sensu*
- 275 Hoff and McDonald, 1980) in some families. Mean percent stem symptoms at 1.42 years
- 276 post-inoculation varied among families ranging from 45.71% to 100% with an overall mean
- of 88.69%, and nine of the 40 families having 100%. The number of stem symptoms
- averaged 9.27 over all families, with family means varying from 1.41 to 17.52. On a
- 279 population basis, families from Lincoln R stands had a lower percentage of stem symptoms
- 280 (86.07%) at the end of the trial (7.42 years post-inoculation) and lower mean number of stem
- 281 symptoms per tree at  $2^{nd}$  assessment (6.38) compared to the random tree selections from
- stands with no rust presence in Cibola (11.63), Santa Fe (11.19) and the susceptible controls
- 283 in Lincoln S (11.36).
- At final inspection (February 2018, 7.42 years post-inoculation) 24.49% of trees had survived
- (Figure 2). There was a total of 446 living individuals and 130 of them were stem symptom
- free, and of the 316 with stem symptoms, only 24 have a severity rating >4 and a number of
- those appeared to have inactive cankers. Family SF-10 had 38.18% survival and 100% of
- surviving trees had stem symptoms present at early assessments but generally low severity.
- Putatively resistant families in Lincoln R had higher survival and ranged from 0% to 84.62%
  (45.00%), while families in Cibola, Santa Fe, and Lincoln S ranged in survival from 0% to
  44% (9.33%).
- 292
- 293 Infection Development
- Across all families 92.68% of seedlings eventually developed stem symptoms. Several
- families experienced a delay in stem symptom development (lower %ESS<sub>2\_4</sub>). In general, the degree of ESS<sub>2\_4</sub> was high 96.39 (table 3) with a range of 84.31-100%. There was a significant
- and negative relationship between the %Survival and %ESS<sub>2-4</sub> (DF = 1,38,  $R^2 = 0.204$ , F = 11.0,
- P < 0.01 (Figure 3A). There was also a significant difference in %ESS<sub>2.4</sub> between Lincoln R
- and the other three populations (DF = 1,38,  $R^2 = 0.40$ , F = 19.25, P < 0.01) (Figure 3B). When
- 300 Lincoln R was removed from the analysis there is still a significant difference of means
- 301 between Lincoln S with both Cibola (P = 0.008) and Santa Fe National Forests (P = 0.005) for
- 302 %ESS<sub>2\_4</sub> (full ANOVA: DF = 2,20,  $R^2 = 0.0.35$ , F = 6.98, P < 0.005) but no significant difference
- between the Cibola and Santa Fe forests (P = 0.94) based on Tukey's HSD post hoc test.
- 304 Only one family (CIF-401) had no survival 2.33 years post-inoculation (3<sup>rd</sup> assessment), but
- 305 six other families had very low survival at this early stage, approximately 5% survival all
- 306 within Cibola or Santa Fe NFs (supplemental table S2). Across all families 67.14% of
- 307 seedlings died from rust by 2.33 years post inoculation (Figure 2). By 3.83 years post
- 308 inoculation (5<sup>th</sup> assessment) mortality was 73.88% and by the final assessment mortality
- 309 reached 75.51% (Figure 2). The six families with the highest survival (> 60%) were all from
- Lincoln R. One family from both the Santa Fe NF (SF-10) and Cibola NF (CIF-404) each had
- 311 moderately high rust survival at 38.18% and 44.19% respectively despite having 100% and
- 312 83% stem symptoms at 1.42 years post inoculation.

- 313 Survival tracked severity across inspections with an overall severity mean of 7.05 (Table 3)
- and a family range of 1.77 to 9. Lincoln R families had significantly lower mean severity
- ratings than the other sites at 5.36 (DF = 1,38,  $R^2$  = 0.49, F = 39.26, P < 0.01). Both Cibola NF
- and Santa Fe NF had one family each with a notable lower mean severity than the other
- 317 nine families in their populations (Figure 2). Trees alive with stem symptoms (SSALV<sub>7</sub>) at
- 318 final assessment (families that only included stem symptom free or 100% mortality were
- removed) had a mean severity rating of 1.48 with a range of 1 to 8 (Figure 4A). This is
- notable as survivors with low severity stem symptoms have little chance of dying later and
- 321 in many cases stem symptoms were bark reactions or inactive cankers. For SSALV<sub>7</sub>
- individuals, there was no significant difference of means between populations for Sev
- rating at 95% confidence. However, several families did differ significantly between eachother for severity (Figure 4B).
- 325
- 326 Pattern of Genetic Resistance

327 There was a significant difference in the mean number of stem symptoms (DF = 3,36,  $R^2$  =

328 0.52 F = 13.05, P < 0.001) among the sampled forests (Supplemental Data Table S3). Lincoln

- R a had much lower average number of stem symptoms at  $2^{nd}$  assessment (12.59) relative to
- Cibola, Lincoln S, and Santa Fe NF (22.39). When Lincoln R families were excluded from
  analysis there was no significant difference between forests and the mean number of stem
- 332 symptoms (Figure 5A).
- 333 Trees that survived the duration of the trial with stem symptoms (n = 316 SSALV<sub>7</sub>) had
- 334 significantly fewer stem symptoms at  $2^{nd}$  assessment (DF = 1,1741,  $R^2$  = 0.16, F = 329.6, P <
- 335 0.001) compared to those that eventually died from blister rust. The mean number of stem
- 336 symptoms at 2<sup>nd</sup> assessment in trees that ultimately survived (SSALV<sub>7</sub>) was 3.32 compared

to those that died from rust 11.39 at final assessment (Figure 5B). The overall mean number

- 338 of stem symptoms for all families at 2<sup>nd</sup> assessment was 9.27 (Figure 5C).
- 339 Trees from Lincoln R and Lincoln S were significantly taller 1.42 years post-inoculation (3<sup>rd</sup>
- 340 year height) (F value = 237.06, DF = 1,1868, P < 0.001) than those found in Cibola and Santa
- 341 Fe (both sites shorter on average by approximately 8cm and 3cm, respectively). Lincoln S
- 342 seedlings were shorter than Lincoln R by approximately 5cm.
- 343 Linear mixed effects models found that on an individual tree basis (parent trees nested
- 344 within forests) the probability of dying from blister rust was positively and statistically
- associated with the number of stem symptoms at  $2^{nd}$  assessment (t-value = 18.95, P < 0.001).
- Additionally, there was a significant relationship between the number of normal cankers
- 347 (nc2) and  $2^{nd}$  year (pre-inoculation) height (*t value* = 6.78, P < 0.001). When individuals from
- 348 the Lincoln NF were removed there was still a positive relationship between nc2 and  $2^{nd}$
- 349 year height (*t value* = 5.69, *P* < 0.001). The number of normal cankers at  $2^{nd}$  assessment was
- positively associated with the number of spots at  $1^{st}$  assessment (*t value* = 5.84, *P* < 0.001) as
- 351 was the overall number of stem symptoms at  $2^{nd}$  assessment (*t value* = 6.01, *P* < 0.001). Dying

- 352 from blister rust was negatively associated with the number of complete bark reactions at
- $2^{nd}$  assessment (*t-value* = -7.834, *P* < 0.001). There was no significant association with the
- number of partial bark reactions at the same assessment (*t value* = -1.798, *P* =0.07). At the
- final assessment (7.42 years post-inoculation) there was a negative significant association
- with the number of complete bark reactions (t *value* = -2.086, *P* = 0.037), and total bark
- 357 reactions (BRall) (t *value* = -15.465, *P* < 0.001).
- 358 As expected, percent survival in a family is significantly and negatively associated with
- percent of seedlings in the family with stem symptoms (*t-value* = -8.448, *P* < 0.001).
- 360 However, there is quite a bit of variation with some families exhibiting a high percentage of
- 361 stem symptoms at 2<sup>nd</sup> assessment but lower final percent rust mortality (Figure 6A). On a
- 362 family basis there was a significant effect of percent bark reactions (BRall) on percent
- 363 survival ( $DF = 1,38, R^2 = 0.45, F \text{ value} = 115.6, P < 0.001$ ) (Figure 6B). At the final assessment
- there was a negative relationship between BR<sub>all</sub> and normal cankers at  $2^{nd}$  assessment (*t*
- 365 value = -6.918, P < 0.001). The appearance of BR<sub>all</sub> culminated around the 5<sup>th</sup> assessment (3.83
- 366 years post-inoculation) (Figure 7).
- 367

368

### MGR vs QR

369 Exact binomial tests of Mendelian segregation ratios 1:1 (Rr x rr) and 3:1 (Rr x Rr) for SSf:SS 370 seedlings identified two families (Lincoln R Id 27 and 79) that failed to differ significantly 371 from 1:1 and there were no families that failed to differ significantly from 3:1(Supplemental 372 table S4). The two potential MGR families (Rr x Rr or Rr x rr) occurred in the Lincoln R 373 stand. However, the range of stem symptom free trees in the 17 Lincoln R parents is 374 relatively continuous and the families also generally have moderate to high bark reactions 375 (BRall7), suggesting the two families that are candidates for MGR are more likely at the 376 upper spectrum in this population for QR. Support for QR is provided by an earlier test where the families included in this trial had previously been tested for MGR and were 377 378 presumed not to carry the Cr3 allele (did not segregate 1:1 in that test). High levels of QR 379 could either mask MGR or express similar to MGR, and the two resistance types may occur 380 in combination in some families since MGR is known to be present at low frequency in this 381 stand.

382

# 383 4 DISCUSSION

384

# 385 *Quantitative disease Resistance in Pinus strobiformis*

386 Many tree species in North America are susceptible to non-native pathogens and pests

- 387 (National Academies of Sciences, Engineering, and Medicine, 2019). The level of
- 388 susceptibility can be extremely high, with local extirpation to potential species level
- 389 extinction possible. The classic forest example is of the American chestnut (*Castanea dentata*)

390 which has been functionally extirpated on the landscape due to the occurrence of two 391 diseases, a root rot and the chestnut blight, but other pests and pathogens are increasing 392 and are greatly impacting forests across much of North America (Fei et al., 2019). Several successful programs are in place to identify the type and frequency of resistance to disease 393 394 and produce seed from resistant populations to aid in restoration or reforestation (Sniezko 395 and Koch, 2017). These programs must first identify the type(s) of genetic resistance, second 396 determine whether to enhance the level of resistance through tree breeding so that it is 397 more usable and lastly determine its durability through field validation trials (Sniezko et al., 398 2020). Trees tend to be long-lived, and because of this resistance must persist for decades or 399 centuries. Although there are different types of resistance to white pine blister rust, 400 including MGR, the most durable type looks to be QR due to the reduced likelihood of C. 401 *ribicola* developing virulence to it (Sniezko et al., 2020). Identifying QR can be challenging 402 because of the time and costs associated with screening seedling families for disease 403 symptoms and discerning the different phenotypes of the seedlings. As further advances in 404 biotechnologies progress, it may be possible in the future to accelerate the process of 405 resistance detection by combining QR trials with genomics (Weiss et al., 2020; Liu et al., 406 2021), genetic field detection (Aglietti et al., 2021) and phenomics (Conrad et al., 2020; 407 Haagsma et al., 2020). Then, field validation trials under natural rust hazard must be 408 installed to better characterize durability and stability of resistance to ensure healthy forests 409 for the long-term following restoration or reforestation with the resistant populations 410 (Sniezko and Koch, 2017). Field validation is one of the best ways to assess whether 411 identified QR in seedlings is representative in adult trees. 412 Most QR screening trials for resistance to white pine blister rust typically last from three to

five years. However, there is a risk of overestimating survival because some families may 413 414 exhibit slow rusting and delayed symptom development which ultimately results in 415 seedling mortality (Sniezko et al., 2011). In this study we have monitored infected P. 416 strobiformis seedlings for 7.5 years post-inoculation, providing higher temporal resolution 417 enabling us to be more confident in our early characterization of QR and associated traits in 418 the species, and that in the most resistant families, individuals may survive with non-lethal 419 stem infections. Additional observations in August 2021 of the trees remaining in the trial 420 (after a thinning) indicate that the resistant trees with stem symptoms survive at least 12 421 years post inoculation (Sniezko, unpublished). Our analysis suggests that there is a usable 422 level of QR in *P. strobiformis*, but that there is only a low frequency, ~10 percent, of parent 423 trees that will have a moderate level of high resistant (surviving) progeny in natural 424 populations. As with most white pine species native to North America, susceptibility of the 425 species in native forests can be very high, for example, greater than 90% of trees cankered. 426 The families with the most effective QR showed levels of survival ranging from 1 to 85%. 427 The Lincoln R population averaged 45% survival, which is the among the highest of any 428 population with QR currently reported for a North American species tested at the Dorena 429 GRC, and very similar to results of testing of canker-free parent trees of P. albicaulis that 430 were selected in stands with 90 percent blister rust mortality (Hoff et al., 2001).

- 431 Additionally, the level of survival (>70%) in the top *P. strobiformis* families far exceeds what
- 432 has been found in much more extensive screening of >4000 seedling families over many
- 433 decades in both *P. monticola* or *P. lambertiana* at Dorena GRC and is similar to some of the
- 434 advance-generation families of *P. monticola* from the breeding program. Trials of wild open
- 435 pollinated *P. monticola* have recorded survival between 3.4% and 9% while *P. lambertiana*
- 436 was noted as having survival between 1.6% and 14% (Kegley and Sniezko, 2004).

From the samples in this study, parent trees with a moderate level of survival and QR
appear, at a frequency of approximately 10% in this portion of the species range. This
frequency stems primarily from the assessment of the Santa Fe and Cibola NF sites which,

- 440 at the time of cone collection, had little rust present and represented random genotype
- selections compared to cone collections in the high infection Bradford Canyon population in
- the Lincoln NF. On balance, the Lincoln R families provide a better look at the range of
- 443 potential levels of QR. In the Lincoln NF near the Bradford Canyon site 85-90% of trees are
- cankered. If we assume that the cankered trees are susceptible then we can begin to see the
- range of QR based on our results. Moreover, because the families from Lincoln R were
- 446 canker-free trees in a stand with ~ 90% susceptibility we can extrapolate overall frequency
- of QR to about 10% of the stand but noticing that the level of QR varied substantially
- 448 among the Lincoln R parents with 15 of 17 parents the survival ranged from 9.5 to 84.6%. It
- 449 can also be difficult to identify bark reactions in the field, so some parent tree selections
- 450 may have had stem symptoms in the past but appeared canker-free when cones were
- 451 selected. The Lincoln S families were cankered trees that were selected as susceptible452 controls. However, even the Lincoln S trees tended to fare slightly better in terms of the
- 452 controls. However, even the Lincoln S trees tended to fare slightly better in terms of the
- 453 production of bark reactions. This is probably due to pollen gene flow of resistant genes
- 454 from the trees with QR in the vicinity.
- 455 When assessing the Lincoln NF families, we find an overall higher level of QR. These
- findings are supported by an earlier smaller trial of *P. strobiformis* from the same Bradford
  Canyon collection site where families had only 45.83% mortality after nearly 14 years of
- 458 monitoring (unpublished data R. Sniezko; Sniezko et al., 2008). Since both MGR and QR
- 459 have now been documented in Bradford Canyon of the Lincoln NF there is potential for at
- 460 least some of the SS-free trees to be the result of pollen from MGR parents however, this is
- 461 expected to be low because 1.) there is a low frequency of MGR; and 2.) greater than 85% of
- 462 trees alive in Bradford Canyon (Lincoln R) have stem symptoms, which also contribute to
- the pollen cloud.
- 464
- 465

### Survival and Slow Fungus Growth/Slow Canker Growth

466 Interestingly, seedling mortality in Cibola and Santa Fe families peaked about two years467 post-inoculation with very little additional mortality occurring at subsequent assessments.

467 post-moculation with very fittle additional mortality occurring at subsequent assessments. 468 In contrast, the Lincoln R families experience more rust mortality at later stages with peak

- 469 mortality occurring around four years post-inoculation. The trees from Bradford Canyon
- 407 mortancy occurring around four years post-moculation. The trees from bradford Canyon
- 470 were on average taller than trees from seed collected in the Cibola and Santa Fe so it might

- 471 take longer to die, or the resistance may be higher (e.g. fewer stem symptoms per tree).
- 472 Mortality in Lincoln R also occurred at lower frequencies and with higher between family
- 473 variation. Survival, however, did decrease slowly over the intervening years (Figure 2A).
- 474 Despite the strong selection pressure in Lincoln R due to the high rust hazard, two of the 17
- families from this collection were among the most susceptible in the trial. The two seed
- parents are approximately 61 meters apart and are on the eastern edge of the sampled
   population. It's possible that they are not receiving as much pollen flow from the QR trees
- 477 population. It's possible that they are not receiving as much pollen flow from the QR trees478 in the vicinity. Other possibilities for their low survival include 1.) they could be in a
- 479 microenvironment of lower infection and thus 'escapes', 2.) show ontogenetic resistance
- 480 that would not be conveyed to young progeny, or 3.) be homozygous for a recessive gene
- 481 for resistance. We suspect they are likely escapes but further investigation would be needed
- 482 to resolve their status.

483 Two families in the trial had percent stem symptoms consistent with 1:1 segregation 484 expected of MGR parent trees. Yet, based on previous testing, the parent trees from Lincoln 485 NF whose progeny were tested in our study were not MGR candidates, however, at least a 486 low frequency of the canker-free seedling may be the result of pollen contribution from the 487 low frequency of MGR parents in the stand, as noted in a small earlier trial (Sniezko et al., 488 2008). Thus, surviving seedlings in these families may be a mix of QR, MGR, and QR + 489 MGR genotypes. Approximately 63% of surviving seedlings had stem symptom at the end 490 of the trial and, in ten families, 100% of living seedlings had stem symptoms. The two 491 parents from Bradford Canyon with relatively low percent stem symptoms (< 65%) are 492 likely near the top of the continuum for QR which can provide resistance at levels like 493 MGR. It is still unclear if the Bradford Canyon location is a hot spot for genetic resistance 494 (not likely), or if it only represents a high hazard site where all the susceptible trees are 495 infected, and the efficacy for resistance selection is greatly enhance by focusing on canker-496 free parent trees which has been noted in trials of *P. monticola* (Kinloch et al., 1999) and *P.* 497 albicaulis (Hoff et al., 2001). It is notable that both MGR and high level of QR have now been 498 documented in the Bradford Canyon stand (Sniezko et al., 2008; Sniezko et al., 2011), and if 499 so, some naturally pyramiding of resistance may already be present. The low degree of QR 500 in the susceptible control families selected from Bradford Canyon, Lincoln S, matched that of the Cibola NF families and were somewhat better performers than the Santa Fe NF 501 families which provides additional support for the likely frequency of QR that can be 502 503 expected across the range of the species, however, this still must be verified.

504

# 505 Bark Reactions and Normal Cankers

506 The necrotic bark reaction allows a tree to develop a wound-periderm on the branch or

stem of the tree that can stop the spread of *C. ribicola* (Struckmeyer and Riker, 1951). Bark

508 reactions in *P. monticola* have exhibited varying effectiveness, where some bark reactions

- stop the growth of the fungus quickly, while in other cases, bark reactions are partial or
- 510 incomplete, and the fungus returns and continues to expand (Hoff, 1986) escaping the tree's

- 511 defenses. Either way, the occurrence of a bark reaction suggests a trait capable of slowing
- 512 the spread of rust and at best stopping the spread of the fungus. Bark reactions have been
- 513 documented in several white pine species, including *P. strobus* (Struckmeyer and Riker,
- 514 1951), P. lambertiana (Kinloch and Littlefield, 1977; Kegley and Sniezko, 2004), and P.
- 515 monticoloa (Hoff, 1986; Kegley and Sniezko, 2004; Sniezko et al., 2014). An inoculated
- 516 seedling can have from 0 to dozens of stem infections, so even when bark reactions do
- 517 occur, they can appear in conjunction with normal cankers on the same tree. The incidence
- of bark reactions is positively associated with increased survival. We have shown here that
- 519 the occurrence of bark reactions appears along a continuum that is associated with QR.
- 520 Complete bark reactions were notable for this species in this trial, and many occurred early
- and were no longer visible by the final assessment or had a very low severity rating of 1 or2.
- 523

## 524 *Quantitative disease Resistance in other White Pines*

525 In this study we equate QR with overall survival. However, we also note that some further 526 QR variation may exist among trees that did not survive. Compared with other five-needle

527 pine species such as *P. monticola* and *P. lambertiana*, *P. strobiformis* has a higher frequency of

- 528 QR, though still low (~10%). Previous studies in *P. monticola* found that both the number of
- 529 bark reactions and percent of a family with bark reactions is significantly and negatively
- 530 correlated with percent stem symptoms as well as the percent of trees that are actively
- 531 infected and alive (SSALV) (Kolpak et al., 2008; Sniezko et al., 2020).

532 Variation in patterns of genetic resistance to white pine blister rust has been addressed in 533 many of the white pine species. In particular, P. monticola, P. flexilis, and P. lambertiana have 534 received a great deal of focus because they each carry a *Cr* allele for MGR. For example, in 535 P. lambertiana MGR resistance was identified (Kinloch et al., 1970; Devey et al., 1995) at low 536 frequencies across its range, typically less than 1% (Kinloch and Dupper, 1987; Kinloch, 537 1992; Kinloch et al., 2018). Similarly, P. monticola exhibits a HR-like phenotype in MGR trees 538 and low frequency (0-8%) of resistance in natural populations though frequencies vary 539 across its range (Kinloch et al., 1999; Kinloch et al., 2003). The overall frequency of MGR 540 resistance in P. flexilis was higher than P. monticola or P. lambertiana, typically near 5% and 541 as high as 14% in the portions of the range tested (Schoettle et al., 2013), and examination of 542 much more of the geographic range is underway. Kinloch and Dupper (1987) reported an 543 HR-like reaction on needles of young P. strobiformis, but similar to P. flexilis, subsequent trials with P. strobiformis have shown HR-like reactions to be less consistent and the use of 544 545 needle phenotypes is often more difficult to utilize in identification of MGR and the 546 presences of a stem symptom-free phenotype may prove to be a better trait for MGR 547 characterization. Because stem symptom free phenotypes can occur in both MGR and QR progenies, molecular markers would be useful to distinguish the underlying type of 548 549 resistance and its control as well or the use of virulent strains of rust (which are currently

550 used in *P. monticola* and *P. lambertiana*).

- 551 An additional consideration for *P. strobiformis* and for the generalization of levels of genetic
- 552 resistance across its range involves the position of its northern range within a moving
- 553 hybrid zone with *P. flexilis*. Recent studies have found that the two species hybridize
- 554 (Menon et al., 2018) and because *P. flexilis* is also known to have low levels of MGR
- 555 (Schoettle et al., 2013), it is unclear if the somewhat higher level of resistance in *P*.
- *strobiformis,* relative to other white pines, is the product of introgression within this hybrid
- 557 zone. One of the results of Menon et al. (2018) was that there was little ongoing gene flow
- 558 between the periphery and core of the *P. strobiformis* range. The question then begs to be
- asked, will much of the species range outside of this hybrid zone be more susceptible to the
- **560** advance of *C. ribicola* in the future? If, however, resistance is exapted (*sensu* Gould and Vrba
- 561 1982) and has evolved in response to a different abiotic or biotic selection pressures, then
- 562 we would hypothesis a more equitable distribution across its range. Efforts have begun to
- 563 further characterize a range-wide baseline frequency and geographic pattern of genetic
- resistance to white pine blister rust in *P. strobiformis*, as well as identifying genomic controls
- 565 of resistance, its effects on the host physiology, and its future distribution under climate 566 change.
- 567 In this study we identified and characterized QR traits in *P. strobiformis* from a portion of its
- 568 northern range limit. In families with highest resistance (generally highest survival), we
- 569 identified 1.) fewer early stem symptoms and a lower frequency of early stem symptoms
- 570 (%ESS<sub>2\_4</sub>), suggesting a slowing of fungal growth, 2.) a higher frequencies of bark reactions
- and 3.) lower severity of infections over the duration of the trial.
- 572 The levels and frequency of QR found here are encouraging, and with a focused selection 573 program more resistant parents can be identified. Field trials to validate the resistance from 574 seedling trials have begun and grafting of resistant parents (or forward selections from 575 resistant families) for placement into a clone bank or orchards have also been begun. The 576 results from this trial and ensuing trials will provide land managers the first source of white
- 577 pine blister rust resistant seed to use in reforestation efforts, but additional testing is needed
- to identify more resistant parents to ensure seedlots used for reforestation are genetically
- 579 diverse. Periodic checks of the resistant parent trees on the Lincoln NF have shown no
- 580 infections, providing cautious optimism that the resistance will be durable.
- 581

# 582 ACKNOWLEDGMENTS

- 583 We would like to thank the technicians and staff at Dorena Genetics Resource Center for
- 584 their dedication and efforts in conducting the trial inspections over the many years,
- 585 particularly the lead efforts of Bob Danchok and Angelia Kegley. We would also like to
- acknowledge the contributions of our late colleague Douglas Savin. JSJ was supported by
- 587 NSF 1340852. Dave Conklin, USFS R3 FHP coordinated the cone collections with others in
- 588 Region 3, and Region 3 FHP provided funding for the study. The manuscript was improved
- 589 thanks to the comments and suggestions from two reviewers.

590

## 591 AUTHOR CONTRIBUTION

- 592 JSJ analyzed and interpreted the data, and led the writing of the manuscript, RAS conceived
- of the paper and assisted with writing, analysis, and interpretation. Both authors reviewed
- and edited the manuscript and approved of the final version.



### 595 **REFERENCES**

- Aglietti, C., Meinecke, C.D., Ghelardini, L., Barnes, I., van der Nest, A., and Villari, C. (2021). Rapid
  detection of pine pathogens *Lecanosticta acicola*, *Dothistroma pini* and *D. septosporum* on
  needles by probe-pased LAMP assays. *Forests* 12(4), 479.
- Bartholomé, J., Brachi, B., Marçais, B., Mougou-Hamdane, A., Bodénès, C., Plomion, C., et al. (2020).
  The genetics of exapted resistance to two exotic pathogens in pedunculate oak. *New Phytologist* 226(4), 1088-1103. doi: 10.1111/nph.16319.
- Bucholz, E.R., Waring, K.M., Kolb, T.E., Swenson, J.K., and Whipple, A.V. (2020). Water relations
  and drought response of *Pinus strobiformis*. *Canadian Journal of Forest Research* 50(9), 905-916.
  doi: 10.1139/cjfr-2019-0423.
- 605 Conklin, D.A. (2004). "Development of the white pine blister rust outbreak in New Mexico". (USDA
   606 Forest Service, Souwestern Region, R3, R3-04-01).
- 607 Conklin, D.A., Fairweather, M.L., Ryerson, D.E., Geils, B.W., and Vogler, D.R. (2009). "White pines,
  608 blister rust, and management in the Southwest". (USDA Forest Service Southwest Region,
  609 R3, R3-FH-09-01).
- 610 Conrad, A.O., Villari, C., Sherwood, P., and Bonello, P. (2020). Phenotyping Austrian pine for
  611 resistance using fourier-transform infrared spectroscopy. *Arboriculture & Urban Forestry*612 46(4), 276-286. doi: 10.48044/jauf.2020.020.
- Devey, M.E., Delfino-Mix, A., Kinloch, B.B., and Neale, D.B. (1995). Random amplified polymorphic
   DNA markers tightly linked to a gene for resistance to white pine blister rust in sugar pine.
   *Proceedings of the National Academy of Sciences* 92(6), 2066-2070.
- Fei, S., Morin, R.S., Oswalt, C.M., and Liebhold, A.M. (2019). Biomass losses resulting from insect
  and disease invasions in US forests. *Proceedings of the National Academy of Sciences* 116(35),
  17371. doi: 10.1073/pnas.1820601116.
- Fins, L., Byler, J., Ferguson, D., Harvey, A., Mahalovich, M.F., McDonald, G., et al. (2002). Return of
  the giants: Restoring western white wine to the Inland Northwest. *Journal of Forestry* 100(4),
  20-26. doi: 10.1093/jof/100.4.20.
- Goodrich, B.A., and Waring, K.M. (2017). *Pinus strobiformis* seedling growth in southwestern US
   mixed conifer forests in managed and non-managed stands. *Forestry: An International Journal of Forest Research* 90(3), 393-403. doi: 10.1093/forestry/cpw057.
- Gould, S.J., and Vrba, E.S. (1982). Exaptation A missing term in the science of form. *Paleobiology*8(1), 4-15. doi: 10.1017/S0094837300004310.
- Haagsma, M., Page, G.F.M., Johnson, J.S., Still, C., Waring, K.M., Sniezko, R.A., et al. (2020). Using
   hyperspectral imagery to detect an invasive fungal pathogen and symptom severity in *Pinus strobiformis* seedlings of different genotypes. *Remote Sensing* 12(24). doi: 10.3390/rs12244041.
- Hawksworth, F.G. (1990). White pine blister rust in southern New Mexico. *Plant Disease* 74(11), 938.
  doi: 10.1094/pd-74-0938a.
- Hoff, R., Bingham, R.T., and McDonald, G.I. (1980). Relative blister rust resistance of white pines.
   *European Journal of Forest Pathology* 10(5), 307-316. doi: 10.1111/j.1439-0329.1980.tb00042.x.
- Hoff, R.J. (1986). Inheritance of the bark reaction mechanism in *Pinus monticola* infected by
   *Cronartium ribicola*. USDA Forest Service, Intermount Research Station, Research Note INT-361.
- Hoff, R.J. (1988). "Blister rust resistance in western white pine for eastern Washington, Idaho, and
  western Montana.," in *Proc. of a western white pine management symposium*, ed. R.S. Hunt.
- 638 (Victoria, BC: Pacific Forestry Centre), 12-20.

639	Hoff, R.J., Ferguson, D., McDonald, G.I., and Keane, R.E. (2001). "Strategies for managing whitebark
640	pine in the presence of white pine blister rust " in Whitebark Pine Communities: Ecology and
641	Restoration, eds. D. Tomback, S.F. Arno & R.E. Keane. (Washington: Island Press), 346-366.
642	Hoff, R.J., and McDonald, G.I. (1980). Resistance to Cronartium ribicola in Pinus monticola: reduced
643	needle-spot frequency. Canadian Journal of Botany 58(5), 574-577. doi: 10.1139/b80-071.
644	Hunt, R.S. (1997). Relative value of slow-canker growth and bark reactions as resistance responses to
645	white pine blister rust. Canadian Journal of Plant Pathology 19(4), 352-357. doi:
646	10.1080/07060669709501059.
647	Jacobi, W.R., Kearns, H.S.J., Cleaver, C.M., Goodrich, B.A., and Burns, K.S. (2018). Epidemiology of
648	white pine blister rust on limber pine in Colorado and Wyoming. Forest Pathology 48(6),
649	e12465. doi: 10.1111/efp.12465.
650	Kegley, A.J., and Sniezko, R.A. (2004). "Variation in blister rust resistance among 226 Pinus monticola
651	and 217 P. lambertiana seedling families in teh Pacific Noorthwest", in: Breeding and genetic
652	resources of five needle pines: Genetics, breeding and adaptability, Proceedings of the IUFRO 2.02.15
653	Working Party Conference eds. R.A. Sniezko, S. Samman, S.E. Schlarbaum & H.B. Kriebel:
654	USDA Forest Service, Rocky Mountain Research Station RMPS-P-32. Fort Collins, CO.), 209-
655	225.
656	King, J.N., David, A., Noshad, D., and Smith, J. (2010). A review of genetic approaches to the
657	management of blister rust in white pines. Forest Pathology 40(3-4), 292-313. doi:
658	10.1111/j.1439-0329.2010.00659.x.
659	Kinloch, B.B. (1992). Distribution and frequency of a gene for resistance to white pine blister rust in
660	natural populations of sugar pine. Canadian Journal of Botany 70(7), 1319-1323. doi:
661	10.1139/b92-165.
662	Kinloch, B.B. (2003). White pine blister rust in North America: Past and prognosis. <i>Phytopathology</i>
663	93(8), 1044-1047.
664	Kinloch, B.B., Burton, D., Davis, D.A., Westfall, R.D., Dunlap, J., and Vogler, D.R. (2012). "Strong
665	partial resistance to white pine blister rust in sugar pine," in <i>Proceedings of the fourth</i>
666	international workshop on the genetics of host-parasite interactions in forestry: Disease and insect
667	resistance in forest trees., eds. R.A. Sniezko, A.D. Yanchuk, J.T. Kliejunas, K.M. Palmieri, J.M.
668	Alexander & S.J. Frankel. (Albany, CA: Pacific Southwest Research Station, Forest Service,
669	U.S. Department of Agriculture), 80-91.
670	Kinloch, B.B., and Comstock, M. (1981). Race of <i>Cronartium ribicola</i> virulent to major gene resistance
671	in sugar pine. <i>Plant Disease</i> 65(7), 604-605. doi: 10.1094/pd-65-604.
672	Kinloch, B.B., and Dupper, G.E. (1987). Restricted distribution of a virulent race of the white pine
673	blister rust pathogen in the western United States. <i>Canadian Journal of Forest Research</i> 17(5),
674	448-451. doi: 10.1139/x87-077.
675	Kinloch, B.B., and Dupper, G.E. (2002). Genetic specificity in the white pine-blister rust pathosystem.
676	<i>Phytopathology</i> 92(3), 278-280. doi: 10.1094/phyto.2002.92.3.278.
677	Kinloch, B.B., and Littlefield, J.L. (1977). White pine blister rust: hypersensitive resistance in sugar
678	pine. Canadian Journal of Botany 55(9), 1148-1155. doi: 10.1139/b77-133.
679	Kinloch, B.B., Parks, G.K., and Fowler, C.W. (1970). White pine blister rust: Simply inherited
680	resistance in sugar pine. <i>Science</i> 167(3915), 193-195.
681	Kinloch, B.B., Sniezko, R.A., Barnes, G.D., and Greathouse, T.E. (1999). A major gene for resistance
682	to white pine blister rust in western white pine from the western Cascade range.
683	<i>Phytopathology</i> 89(10), 861-867. doi: 10.1094/phyto.1999.89.10.861.

- Kinloch, B.B., Sniezko, R.A., and Dupper, G.E. (2003). Origin and distribution of *Cr2*, a gene for
  resistance to white pine blister rust in natural populations of western white pine. *Phytopathology*® 93(6), 691-694. doi: 10.1094/PHYTO.2003.93.6.691.
- Kinloch, B.B., Sniezko, R.A., and Dupper, G.E. (2004). Virulence gene distribution and dynamics of
  the white pine blister rust pathogen in western North America. *Phytopathology* 94, 751-758.
- Kinloch, B.B., Sniezko, R.A., Savin, D.P., Danchok, R., Kegley, A., Burton, D., et al. (2018). "Patterns
  of variation in blister rust resistance in sugar pine (*Pinus lambertiana*)," in *Proceedings of the IUFRO joint conference: Genetics of five-needle pines, rusts of forest trees, and Strobusphere*, eds.
- A.W. Schoettle, R.A. Sniezko & J.T. Kliejunas. (Fort Collins, CO: U.S. Department of
  Agriculture, Forest Service, Rocky Mountain Research Station), 124-128.
- Kolpak, S.E., Sniezko, R.A., and Kegley, A.J. (2008). Rust infection and survival of 49 *pinus monticola* families at a field site six years after planting. *Annals of Forest Research* 51, 67-80.
- Liu, J.-J., Fernandes, H., Zamany, A., Sikorski, M., Jaskolski, M., and Sniezko, R.A. (2021). In-vitro
  anti-fungal assay and association analysis reveal a role for the *Pinus monticola* PR10 gene
  (PmPR10-3.1) in quantitative disease resistance to white pine blister rust. *Genome* 64(7), 693704. doi: 10.1139/gen-2020-0080.
- Liu, J.-J., Hammett, C., and Sniezko, R.A. (2013). *Pinus monticola* pathogenesis-related gene PmPR102 alleles as defense candidates for stem quantitative disease resistance against white pine
  blister rust (*Cronartium ribicola*). *Tree Genetics & Genomes* 9(2), 397-408. doi: 10.1007/s11295012-0561-0.
- Liu, J.-J., Williams, H., Zamany, A., Li, X.-R., Gellner, S., and Sniezko, R.A. (2020). Development and
  application of marker-assisted selection (MAS) tools for breeding of western white pine
  (*Pinus monticola* Douglas ex D. Don) resistance to blister rust (*Cronartium ribicola* J.C. Fisch.)
  in British Columbia. *Canadian Journal of Plant Pathology* 42(2), 250-259. doi:
  10.1080/07060661.2019.1638454.
- Looney, C.E., and Waring, K.M. (2012). Patterns of forest structure, competition and regeneration in
   southwestern white pine (*Pinus strobiformis*) forests. *Forest Ecology and Management* 286, 159 170. doi: https://doi.org/10.1016/j.foreco.2012.09.008.
- Menon, M., Bagley, J.C., Friedline, C.J., Whipple, A.V., Schoettle, A.W., Leal-Sàenz, A., et al. (2018).
  The role of hybridization during ecological divergence of southwestern white pine (*Pinus strobiformis*) and limber pine (*P. flexilis*). *Molecular Ecology* 27(5), 1245-1260. doi:
  10.1111/mec.14505.
- Menon, M., Bagley, J.C., Page, G.F.M., Whipple, A.V., Schoettle, A.W., Still, C.J., et al. (2021).
  Adaptive evolution in a conifer hybrid zone is driven by a mosaic of recently introgressed and background genetic variants. *Communications Biology* 4(1), 160. doi: 10.1038/s42003-020-01632-7.
- Menon, M., Landguth, E., Leal-Saenz, A., Bagley, J.C., Schoettle, A.W., Wehenkel, C., et al. (2020).
  Tracing the footprints of a moving hybrid zone under a demographic history of speciation
  with gene flow. *Evolutionary Applications* 13(1), 195-209. doi: 10.1111/eva.12795.
- National Academies of Sciences, E., and Medicine (2019). "Forest health and biotechnology:
   Possibilities and considerations". (Washington, DC: National Academies Press.).
- RCoreTeam (2020). "R: a language and environment for statistical computing", in: *R Foundation for Statistical Computing*. (Vienna, Austria).
- Schoettle, A.W. (Year). "Ecological roles of five-needle pines in Colorado: Potential consequences of
   their loss", in: *Breeding and Genetic Resources of Five-Needle Pines: Growth, Adaptability, and Pest*

729 Resistance, eds. R.A. Sniezko, S. Samman, S.E. Schlarbaum & H.B. Kriebel: USDA Forest 730 Service, Rocky Mountain Research Station, Fort Collins, CO ), 124-135. 731 Schoettle, A.W., Sniezko, R.A., Kegley, A., and Burns, K.S. (2013). White pine blister rust resistance 732 in limber pine: Evidence for a major gene. *Phytopathology* 104(2), 163-173. doi: 10.1094/phyto-733 04-13-0092-r. 734 Seager, R., and Vecchi, G.A. (2010). Greenhouse warming and the 21st century hydroclimate of 735 southwestern North America. Proceedings of the National Academy of Sciences 107(50), 21277-736 21282. doi: 10.1073/pnas.0910856107. 737 Shirk, A.J., Cushman, S.A., Waring, K.M., Wehenkel, C.A., Leal-Sáenz, A., Toney, C., et al. (2018). 738 Southwestern white pine (*Pinus strobiformis*) species distribution models project a large range 739 shift and contraction due to regional climatic changes. Forest Ecology and Management 411, 740 176-186. doi: https://doi.org/10.1016/j.foreco.2018.01.025. 741 Sniezko, R.A. (2006). Resistance breeding against nonnative pathogens in forest trees - current 742 successes in North America. Canadian Journal of Plant Pathology 28(sup1), S270-S279. doi: 743 10.1080/07060660609507384. 744 Sniezko, R.A., Johnson, J.S., and Savin, D.P. (2020). Assessing the durability, stability, and usability 745 of genetic resistance to a non-native fungal pathogen in two pine species. Plants, People, 746 Planet 2(1), 57-68. doi: DOI: 10.1002/ppp3.49. 747 Sniezko, R.A., Kegley, A., and Danchok, R. (2008). White pine blister rust resistance in North 748 American, Asian and European species - results from artificial inoculation trials in Oregon. 749 Annals of Forest Research 51, 53-66. 750 Sniezko, R.A., and Koch, J. (2017). Breeding trees resistant to insects and diseases: putting theory 751 into application. Biological Invasions 19(11), 3377-3400. doi: 10.1007/s10530-017-1482-5. 752 Sniezko, R.A., Mahalovich, M.F., Schoettle, A.W., and Vogler, D.R. (2011). "Past and current 753 investigations of the genetic resistance to Cronartium ribicola in high-elevation five-needle 754 pines," in The future of hight-elevation, five-needle white pines in western North America. Proc High 755 Five Symp RMRS-P-63, eds. R.E. Keane, D.F. Tomback, M.P. Murray & C.M. Smith. (USDA 756 Forest Service Rocky Mountain research Station, Fort Collins, CO), 246-264. 757 Sniezko, R.A., Smith, J., Liu, J.-J., and Hamelin, R. (2014). Genetic resistance to fusiform rust in 758 southern pines and white pine blister rust in white pines – A contrasting tale of two rust 759 pathosystems – Current status and future prospects. Forests 5(9), 2050-2083. 760 Struckmeyer, B.E., and Riker, A.J. (1951). Wound-periderm formation in white pine trees resistant to 761 blister rust. Phytopathology 41, 276-281. 762 Tomback, D.F., and Achuff, P. (2010). Blister rust and western forest biodiversity: ecology, values 763 and outlook for white pines. Forest Pathology 40(3-4), 186-225. doi: 10.1111/j.1439-764 0329.2010.00655.x. Vázquez-Lobo, A., De La Torre, A.R., Martínez-García, P.J., Vangestel, C., Wegzryn, J.L., Ćalić, I., et 765 766 al. (2017). Finding loci associated to partial resistance to white pine blister rust in sugar pine 767 (Pinus lambertiana Dougl.). Tree Genetics & Genomes 13(5), 108. doi: 10.1007/s11295-017-1190-4. 768 Weiss, M., Sniezko, R.A., Puiu, D., Crepeau, M.W., Stevens, K., Salzberg, S.L., et al. (2020). Genomic 769 basis of white pine blister rust quantitative disease resistance and its relationship with 770 qualitative resistance. The Plant Journal n/a(n/a). doi: 10.1111/tpj.14928. 771 772

### 773 Figure Captions:

### 774 BOX 1: Resistance Continuum in Pinus. strobiformis

775 Genetic resistance to the disease white pine blister rust is generally split between **major** 776 gene resistance (MGR) and quantitative disease resistance (QR). Each type of resistance is 777 associated with different traits and frequencies. Infection of pine hosts with Cronartium 778 ribicola occurs when (A) basidiospores of C. ribicola disperse from the alternate host, mostly 779 notably species of *Ribes*, and enter the stomata of the pine needles. (B) The fungal hyphae 780 spread through the needle tissue and forms needle spots. These needle spots may be 781 diagnostic for MGR if a hypersensitive-like (HR-like) reaction occurs and arrests the further 782 spread of the fungus. This results in trees that are stem symptom free. However, if MGR is 783 not present than the fungal hyphae progress through the plant tissue and usually enter the 784 bole of the tree. (C) If QR is present than several traits may manifest including necrotic bark 785 lesions that wall off the spread of the fungus – known as bark reactions. Additional traits 786 include slow fungal growth, decreased numbers of stem symptoms and decreased severity 787 of infection. (D) If the tree is susceptible than the fungus will form a normal canker disrupt 788 normal vascular processes eventually girdling and killing the tree. Eventually the fungus 789 will proceed to develop aeciospores spores that will erupt from the canker and disperse to

790 reinfect the alternate *Ribes* host.

791 (E) The best methods for identifying and tracking the progression of the disease and the 792 development of different QR traits requires growing seedlings from open-pollinated cones 793 in common gardens (Photo of USDA Forest Service, Dorena Genetic Resource Center, 794 Cottage Grove, OR. (F) As phenotypes develop in the inoculated seedling families a 795 continuous range of variation in traits may become obvious, where, for example, family 1 796 may have a higher number of stem symptoms compared to another family 2. These 797 differences are often associated with QR. (G) The most useful indicator of QR is survival 798 even in the presence of infection. In inoculation trials, susceptible families (orange) will 799 usually reach 100% stem symptoms and 100% mortality quickly compared to QR families 800 (low QR black and high QR dark brown) that might range between 0-100% survival but 801 mortality is delayed or, in the case of top QR families, avoided. The seedlings in the top QR 802 families usually have some seedlings that have no stem symptoms, and some that have bark 803 reactions or slow growing cankers, as well as some seedlings that died from rust infection. 804 MGR, in contrast, is characterized by a family that segregates 1:1 or 3:1 for a SS-Free 805 phenotype and 50% or greater survival (purple), with the progression of the fungus being 806 stopped in the needles. Obvious HR-like spot phenotypes, like those common in P. 807 lambertiana or P. monticola MGR seedlings, may or may not always clearly appear in Pinus 808 strobiformis. Needle shed can also sometimes be notably present in MGR seedling families. 809 MGR, when present, may mask the expression of QR and current resistance trials based on 810 common garden approaches are generally unable to determine if families have both MGR 811 and QR traits, unless a virulent strain of rust is used to overcome MGR.

- 812 **(H)** Progression of the disease following infection of two year old seedlings occurs over
- 813 several years with needle spots first appearing approximately 7-9 months after infection
- 814 with canker and stem symptom formation occurring approximately 12 months after
- 815 infection and continuing with some families having delayed symptom development.
- 816 Mortality follows the expansion of cankers eventually girdling the tree. The timing of
- 817 inspections will attempt to follow the natural progression of the disease.
- 818

nreview

- Figure 1: Open-pollinated cones from 40 trees were selected from three national forests
- 820 (black dots): The Santa Fe in northern New Mexico, Cibola in western New Mexico, and the
- 821 Lincoln in southern New Mexico. These population represent a portion of the disjunct
- 822 northern periphery of the *Pinus strobiformis* range (extending south into Mexico) within the
- 823 range of white pines (green). USDA Forest Service unit boundaries are shown in light gray.
- 824

Figure 2: (A) Percent family survival. Families ranged widely in percent survival with a greater
distribution of survival occurring within Lincoln R (green). Both Cibola NF (pink) and Santa Fe NF

- 827 (purple) each had one family that had moderate survival: 44.19% and 38.18% respectively. (B) Mean
- 828 family severity of infection also had a wide range. Almost a continuous distribution of mean values
- from low (<2.5) to 9 (all trees dead) on the 0 to 9 scale. Note that both Cibola and Santa Fe NF have one family each with lower mean severity.
- 831

Figure 3: %ESS<sub>2\_4</sub>: (A) There is a significant and negative relationship between the %Survival

and  $%ESS_{2_4}$  (y=291.99x-2.78, P < 0.001, r = 0.532). (B) Most families in the Cibola (pink) and

834 Santa Fe (purple) National Forests had 100% stem symptom development at 1.42 years post-

835 inoculation (2<sup>nd</sup> assessment). Both Lincoln R (green) and Lincoln S (blue) had a lower

- 836 %Early Stem Symptom development suggesting a level of QR.
- 837

838 Figure 4: For trees that remained alive (stem symptom free and 100% mortality families

removed) with stem symptoms (SSALV<sub>7</sub>) at the final assessment (7.42 years post-

840 inoculation) the mean severity remained relatively low (1.48). (A) there was no significant

841 difference at 95% confidence in the mean severity for SSALV<sub>7</sub> at the forest population scale.

842 There was a range of severity suggesting that several trees may have quantitative

resistance. **(B)** There were some significant differences between families (P < 0.001 = \*\*\*, P

<0.01 = \*\*, and P < 0.05 = \*). Gray points reflect jittered individual seedling values for severity within forest stands (A) or families (B).

846

Figure 5: (A) Lincoln R (green) had significantly fewer (P < 0.01) stem symptoms than the Cibola

848 (pink), Santa Fe (purple) and Lincoln S (blue) National Forests. \*\*\* = P < 0.01 (B) The difference in

849 mean number of stem symptoms at 2<sup>nd</sup> assessment (1.42 years post-inoculation) between trees that

850 remained alive with stem symptoms (SSALV<sub>7</sub>) at final assessment and those that died of rust.

- 851 SSALV<sub>7</sub> trees had significantly fewer stem symptoms than their counterparts that died of rust (P < P
- 852 0.01). **(C)** The frequency distribution of all stem symptoms (normal cankers + complete bark
- 853 reactions + partial bark reactions) at 2<sup>nd</sup> assessment had a mean number of stem symptoms of 9.27.
- 854
- 855

- 856 Figure 6: **(A)** Final family percent survival is negatively associated with percent stem
- symptoms at  $2^{nd}$  assessment (%SS2) (P < 0.001). Lincoln R (green) had significantly lower
- 858 percent stem symptoms and higher overall survival compared to Lincoln S (blue), Cibola
- 859 (pink) and Santa Fe (purple) National Forests. **(B)** Family percent bark reactions + partial
- bark reactions (BR<sub>all7</sub>) were positively and significantly related to overall survival (P < P
- 861 0.001). Lincoln R had significantly more BRall than either Cibola or Santa Fe forests. Error
- 862 bars +/- SE point size increases with the number of seedlings in a family. Seed families that
- 863 have %SS2 > 80% and % Survival > 25% are labeled in both panels.
- 864
- 865 Figure 7: Temporal trend in family percent complete bark reactions and partial bark
- 866 reactions (%BRal). Lincoln R (green), Lincoln S (blue), Cibola (pink) and Santa Fe (purple)
- 867 %BRall continued to rise throughout the trial with families reaching a maximum mean
- 868 %BRall near 5<sup>th</sup> assessment (3.83 years post-inoculation: grey dashed line), although most
- 869 bark reactions were noted less than two years post-inoculation.
- 870

### 871 Tables

- Table 1: Forty parent trees were selected from three national forests. Lincoln NF included
- 873 three selections that were cankered to serve as susceptible controls (Lincoln S). Collection
- 874 type is classified as random selection if no blister rust was observed in the stand or resistant
- if trees were selected as canker-free from heavily infected stands. General stand location
- 876 and elevation are noted.

National Forest	# of Families	# seedlings	Collection Type	Mean Latitude(dd)	Mean Longitude(dd)	Mean Elevation(m)
Cibola	10	514	Random	35.15698237	-108.1065316	2732
Lincoln	17 3	808 128	Resistant Susceptible	32.97874753	-105.7162916	2701
Santa Fe	10	423	Random	35.96582414	-106.2981034	2656
Total	40	1873				

# 877 878

879 Table 2. Assessment periods and traits measured during inspections at Dorena GRC. Time

880 periods are measured as months post-inoculation (mpi) and years post-inoculation (ypi) in

881 parentheses. The *x*'s indicate the trait measured at a specific assessment. Abbreviations for

882	each trait o	defined in	parentheses	after trait name	<u>.</u>

	Assessment # and approximate months post-inoculation (mpi)											
Trait	0.5	1	2	3	4	5	6	7				
Ifait	7.5 mpi	9 mpi	17 mpi	28 mpi	39.5 mpi	46.5 mpi	62.8 mpi	89.23				
	(0.63)	(0.75)	(1.42)	(2.33)	(3.25)	(3.8)	(4.5)	(7.4)				
Height (YrHT)	х		х									
Spots (#Spots)	х	x	х									
Survival (Rsurv)		x	x	х	х	х	х	x				
Infection severity (Sev)		x	x	х	х	х	х	x				
Total # stem symptoms (#SS)		x	x	х	х	х	х	x				
# bole infections (#BI)			x	х	х	х	х	x				
# partial bark reactions (#PBR)			x	x	x	x	х	x				
# complete bark reactions (#BR,	<u>_</u> )		x	х	х	х	x	x				

### 883 884

004

885

- 886Table 3: Family means for progeny from 40 open pollinated *Pinus strobiformis* trees. TreeID
- 887 corresponds to the seed parent followed by National Forest where the cones were collected.
- 888 The #seedlings are the individuals inoculated in the trial from each family. Traits means
- include year two and year three height (cm), number of spots (#Spot<sub>1</sub>) and percent spots
- 890 (%Spot1) at 1<sup>st</sup> assessment, number of stem symptoms (#SS2), percent stem symptoms (%SS2)
- at 2<sup>nd</sup> assessment. Percent stem symptoms (%SS<sub>7</sub>), percent bark reactions all (partial and
- 892 complete) (%BRall<sup>7</sup>), severity of infection (%Sev<sup>7</sup>), percent of trees alive with stem
- symptoms (%SSALV<sub>7</sub>), percent of stem symptom-free (%SS Free<sub>7</sub>) and percent survival at
- 894 final assessment (%Survival<sup>7</sup>). The composite percent early stem symptoms between 2<sup>nd</sup> and
- <sup>895</sup> 4<sup>th</sup> assessment (ESS<sub>2\_4</sub>). The trial averages are at bottom of the table in Bold. The complete
- table of family means can be found in supplemental table S1.

	National		Mean	Mean	Mean			Mean							Mean		
TreeID	Forest	#seedlings	Yr2Ht (cm)	Yr3Ht (cm)	#Spot <sub>1</sub>	%Spot <sub>1</sub>	%Spot <sub>2</sub>	#SS <sub>2</sub>	%SS₂	%SS <sub>7</sub>	%BR <sub>all7</sub>	%SSF <sub>7</sub>	%SSALV <sub>7</sub>	%RSurv <sub>7</sub>	sev <sub>7</sub>	%ESS <sub>1_4</sub>	%ESS <sub>2_4</sub>
2	Lincoln R	38	21.21	45.95	44.18	100.00	84.62	4.55	63.16	73.68	64.29	26.32	60.00	65.79	3.74	44.44	88.89
11	Lincoln R	56	16.32	31.21	34.84	98.21	58.33	6.86	89.29	96.43	55.56	3.57	89.47	33.93	6.05	73.58	94.34
15	Lincoln R	59	24.74	47.99	35.64	100.00	95.45	7.00	86.44	89.83	60.38	10.17	83.78	62.71	3.83	63.46	98.08
16	Lincoln R	23	16.38	29.43	47.09	100.00	83.33	5.70	73.91	82.61	31.58	17.39	60.00	43.48	5.61	57.89	89.47
27	Lincoln R	39	19.70	42.85	27.49	100.00	60.00	1.41	51.28	64.10	76.00	35.90	57.58	84.62	1.77	45.45	90.91
50	Lincoln R	43	21.69	40.45	61.28	100.00	68.97	8.40	76.74	88.37	52.63	11.63	58.33	27.91	6.95	63.16	86.84
51	Lincoln R	54	21.11	38.56	27.46	100.00	67.86	5.41	77.78	90.74	57.14	9.26	78.26	42.59	6.06	66.67	87.50
64	Lincoln R	57	23.29	40.26	27.58	100.00	79.41	7.23	75.44	85.96	40.82	14.04	63.64	38.60	5.74	55.10	87.76
72	Lincoln R	58	18.40	32.07	20.43	98.28	80.36	11.45	98.28	100.00	20.69	0.00	0.00	0.00	9.00	53.45	98.28
74	Lincoln R	59	22.84	42.86	27.92	96.61	89.47	10.05	100.00	100.00	30.51	0.00	100.00	1.69	8.85	50.85	100.00
79	Lincoln R	35	15.37	34.40	39.26	100.00	66.67	3.11	45.71	51.43	55.56	48.57	34.62	74.29	2.63	47.06	94.12
81	Lincoln R	20	19.47	43.70	42.95	100.00	100.00	5.85	90.00	95.00	73.68	5.00	93.33	75.00	3.20	72.22	100.00
82	Lincoln R	60	21.09	42.60	28.40	100.00	65.79	4.47	75.00	81.67	63.27	18.33	50.00	36.67	5.95	46.94	91.84
83	Lincoln R	58	21.50	37.76	19.95	98.28	80.65	5.48	74.14	86.21	48.00	13.79	70.37	46.55	5.12	55.10	87.76
84	Lincoln R	47	19.67	39.00	33.74	100.00	64.71	5.02	89.36	93.62	63.64	6.38	90.00	63.83	4.02	74.42	97.67
103	Lincoln R	60	21.47	40.55	63.73	100.00	74.07	4.88	71.67	88.33	54.72	11.67	75.86	48.33	5.00	58.82	84.31
104	Lincoln R	42	15.70	30.70	18.64	100.00	93.55	11.62	95.24	95.24	22.50	4.76	75.00	19.05	7.55	67.50	100.00
CIF-401	Cibola	21	17.70	27.75	14.19	100.00	50.00	8.48	100.00	100.00	42.86	0.00	0.00	0.00	9.00	42.86	100.00
CIF-402	Cibola	59	19.42	33.22	25.10	100.00	91.43	10.95	98.31	98.31	3.45	1.69	50.00	3.39	8.71	81.03	100.00
CIF-403	Cibola	50	16.58	26.65	80.60	100.00	83.78	14.68	98.00	100.00	18.00	0.00	100.00	10.00	8.36	63.27	100.00
CIF-404	Cibola	43	17.61	30.36	84.05	100.00	79.17	10.84	83.72	86.05	48.65	13.95	68.42	44.19	5.72	58.33	100.00
CIF-405	Cibola	51	17.62	28.12	60.40	100.00	100.00	13.33	92.16	94.12	16.67	5.88	70.00	19.61	7.59	63.83	100.00
CIF-406	Cibola	51	16.18	24.80	68.27	100.00	87.50	9.71	96.08	98.04	6.00	1.96	50.00	3.92	8.65	54.00	98.00
CIF-407	Cibola	60	20.26	35.28	48.20	100.00	82.22	10.03	95.00	95.00	14.04	5.00	50.00	10.00	8.13	70.18	100.00
CIF-409	Cibola	60	20.07	32.22	55.33	100.00	82.69	12.00	95.00	96.67	8.62	3.33	60.00	8.33	8.25	70.69	98.28
CIF-410	Cibola	60	23.38	37.16	17.18	98.33	75.51	12.07	95.00	100.00	33.33	0.00	100.00	11.67	8.15	60.00	95.00
CIF-411	Cibola	59	22.77	34.52	57.25	100.00	100.00	14.19	94.92	94.92	8.93	5.08	0.00	5.08	8.54	62.50	100.00
SF-1	Santa Fe	42	18.31	30.46	35.26	100.00	87.18	10.07	97.62	97.62	14.63	2.38	0.00	2.38	8.79	56.10	100.00
SF-10	Santa Fe	55	21.54	35.04	42.42	100.00	69.57	9.36	100.00	100.00	52.73	0.00	100.00	38.18	6.11	89.09	100.00
SF-2	Santa Fe	25	16.69	29.08	66.52	100.00	100.00	17.52	100.00	100.00	0.00	0.00	0.00	0.00	9.00	80.00	100.00
SF-3	Santa Fe	50	16.66	26.52	29.66	100.00	69.23	8.80	98.00	100.00	18.00	0.00	100.00	4.00	8.66	52.00	98.00
SF-4	Santa Fe	12	13.08	25.17	43.58	100.00	100.00	10.33	100.00	100.00	0.00	0.00	0.00	0.00	9.00	50.00	100.00
SF-5	Santa Fe	37	13.31	23.15	45.59	100.00	82.86	9.68	100.00	100.00	16.22	0.00	100.00	2.70	8.86	48.65	100.00
SF-6	Santa Fe	43	20.74	34.88	59.91	100.00	72.22	11.79	100.00	100.00	20.93	0.00	100.00	9.30	8.49	76.74	100.00
SF-7	Santa Fe	59	20.56	33.59	31.32	100.00	75.51	13.25	100.00	100.00	16.95	0.00	100.00	1.69	8.95	61.02	100.00
SF-8	Santa Fe	40	17.62	28.33	54.93	100.00	62.50	9.75	100.00	100.00	25.00	0.00	100.00	2.50	8.98	72.50	100.00
SF-9	Santa Fe	60	22.21	36.53	31.20	100.00	83.02	11.38	98.33	100.00	23.33	0.00	100.00	3.33	8.95	63.33	98.33
Susc_1	Lincoln S	47	19.18	39.73	44.67	100.00	84.21	10.34	93.62	95.74	33.33	4.26	66.67	12.77	7.91	63.64	100.00
Susc_2	Lincoln S	48	20.01	33.43	15.54	95.83	83.33	9.81	87.50	93.75	22.22	6.25	50.00	12.50	7.90	64.44	93.33
Susc_3	Lincoln S	33	15.82	28.36	28.21	100.00	86.21	13.94	90.91	93.94	0.00	6.06	33.33	9.09	8.39	61.29	96.77
		1873	19.18	34.37	41.00	99.64	80.03	9.27	88.69	92.68	32.87	7.32	63.47	24.49	7.05	61.54	96.39









Figure 4.JPEG





Figure 5.JPEG





Figure 6.JPEG







![](_page_37_Figure_1.jpeg)