January 7, 2022

Deborah Trent

Linda Jackson, Payette Forest Supervisor Stibnite Gold Project 500N. Mission St., Building 2 McCall, ID. 83638

Ms Jackson.

I am commenting about what appears to be a major shift in direction from the mining of gold, to the mining of Antimony. The recent large sum of money from the US Department of Defense to Perpetua Resources to do an environmental and engineering studies on antimony trisulfide, has me very concerned.

There has been recent research about Antimony in 2021 by a group of International Scientists (see enclosed document: Environment International) that proves that the mining of Antimony is potentially toxic to ecosystems and public health via the accumulation in the food chain. However, there are gaps in understanding how poisonous and carcinogenic it is. Also unclear is what are the mechanisms that cause the toxicity.

Antimony in certain biochemical states is one of the most explosive elements on earth! The transportation, storing, and processing of it is very dangerous. Will it be stored on site? If not where and what kind of transport in and out of the mountains and on our Highways 55 and 95 would permit it? What if there was a fire in the area that would come into contact with it? OR God forbid, an act of terrorism?

If Antimony leaches into the South Fork of the Salmon or Antimony dust particles build up in the soils, how would this affect fish, wildlife, and the entire ecosystem for years to come?

It seems to me that there is no amount of money in the world that would cover all these "what ifs" with a 100% guarantee that some sort of event could be avoided. Any of these situations could happen and the close proximity to our communities and Wild and Scenic Salmon River, would be complete devastation.

The RISKS OUTWEIGH THE BENEFITS here. This is a short term bad decision that could have very irreversible long term damaging consequences.

Please, chose the NO- Action alternative! I do not see any way that the Stibnite Mine should be allowed to proceed.

Sincerely,

Deborah Trent

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Antimony contamination and its risk management in complex environmental settings: A review

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ABSTRACT

Antimony (Sb) is introduced into soils, sediments, and aquatic environments from various sources such as weathering of sulfide ores, leaching of mining wastes, and anthropogenic activities. High Sb concentrations are toxic to ecosystems and potentially to public health via the accumulation in food chain. Although Sb is poisonous and carcinogenic to humans, the exact mechanisms causing toxicity still remain unclear. Most studies concerning the remediation of soils and aquatic environments contaminated with Sb have evaluated various amendments that reduce Sb bioavailability and toxicity. However, there is no comprehensive review on the biogeochemistry and transformation of Sb related to its remediation. Therefore, the present review summarizes: (1) the sources of Sb and its geochemical distribution and speciation in soils and aquatic environments, (2) the biogeochemical processes that govern Sb mobilization, bioavailability, toxicity in soils and aquatic environments, and possible threats to human and ecosystem health, and (3) the approaches used to remediate Sb-contaminated soils and water and mitigate potential environmental and health risks. Knowledge gaps and future research needs also are discussed. The review presents up-to-date knowledge about the fate of Sb in soils and aquatic environments and contributes to an important insight into the environmental hazards of Sb. The findings from the review should help to develop innovative and appropriate technologies for controlling Sb bioavailability and toxicity and

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1. Introduction

Antimony (Sb) is one of the most commonly used metallic element, with global reserve of 1.9 million tons. According to recent report, almost 80% of Sb production was concentrated in China, Russia, and Bolivia (United States Geological Survey, 2021). By 2020, China was the leading producer of Sb worldwide, accounting for nearly 80,000 metric tons, and Russia the second highest (Statista, 2021). Sb is most commonly used as a fire retardant in various products such as toys, carseat covers, light-aircraft-engine covers, and clothing for children and fire fighters (Bagherifam et al., 2019a, Filella et al., 2020, Hu et al., 2021), which accounts for around 60% of global Sb consumption. Sb is also frequently used as a glass decolorizer, a catalyst in plastic production, in the manufacture of paint pigments, and in metal alloys for ammunition and battery production (Diquattro et al., 2021; Guo et al., 2018; Okkenhaug et al., 2016). Sb is released into soils and aquatic environments from several sources, the weathering of sulfide ores, leaching from mining wastes, and anthropogenic activities such as smelting, metallurgical operations, and shooting (Diquattro et al., 2020; Diquattro et al., 2021). For example, in the vicinity of the world's biggest Sb-mining site in Hunan Province, China, Sb concentrations in mining soils range from 101 to 5,045 mg kg⁻¹ (He, 2007), and from 17 to 288 μ g L⁻¹ in water samples (Guo et al., 2018; Wang et al., 2011). In the Extremadura province of Spain, Sb concentrations at three abandoned Sb-mining areas range from 225 to 2,449 mg kg⁻¹ (Murciego acal., 2007). In the United States (U.S), an estimated 1,900 tons of Sb have been released from approximately 3,000 military shooting ranges and 9,000 public shooting ranges (Wan et al., 2013). Shooting-range soils are considered a key anthropogenic source of Sb in several countries (Okkenhaug et al., 2013), notably Australia (Sanderson et al., 2014), Korea (Ahmad et al., 2014), Finland (Sorvari, 2007), Norway (Stromseng et al., 2009), and Canada (Laporte-Saumure et al., 2011).

Similar to arsenic (As), Sb possesses an s²p³ outer orbital electron configuration and thus occurs in the form of (-III, 0, +III, and + V) (Diquattro et al., 2020). In the natural pH range (3-10), the deprotonated state of antimonic acid or Sb(OH)₆⁻ is the predominant Sb(V) species, which is stable over a wide range of soil reduction-oxidation (redox) potential, whereas under anaerobic conditions, Sb(III) occurs as neutral antimonous acid (Sb(OH)₃) (Diquattro et al., 2020; Rinklebe et al., 2020). Sb(OH)₆⁻ has also been found the leading form in polluted sites close to mineral ore smelteries and at shooting ranges (Johnson et al., 2005; Takaoka et al., 2005).

A high Sb concentration in soil or sediments is toxic to ecosystems, and potentially affects human health via accumulation in the food chain (Antoniadis et al., 2019; Shaheen et al., 2019, Shahid et al., 2019). Once entering human bodies by either direct (inhalation or ingestion) or indirect (via the food chain) exposure, Sb and Sb-derived compounds react with sulfhydryls in human tissue, leading to cellular hypoxia by inhibiting enzymatic action and disturbing cellular ionic balance. These adverse impacts eventually result in metabolic malfunctioning and impairment of the nervous system and as vital organs (Yang et al., 2015). Generally, the toxicity of various Sb speciation is in the following order: antimonite (III) > antimonite (V) > organo-antimonials (e.g., methylated species) (Wei et al., 2015). Sb is classed as a priority contaminant by both the US Environmental Protection Agency (USEPA) and the Council of the European Union (EU) (Bagherifam et al., 2019b; Hua et al., 2021). The maximum acceptable values of Sb in consumable water and soil regulated by the World Health Organization (WHO) are 0.020 mg L⁻¹ and 36 mg kg⁻¹, respectively (Chang et al., 2002; Guo et al., 2009).

Human toxicity of Sb(III), a reduced form of Sb, is tenfold higher than

that of the oxidized form (Sb(V)) (Filella et al., 2020). Xue et al. (2017) reported entry of Sb into the human body via ingestion of a contaminated diet and dermal contact, which eventually disturbs the viability, formation, and secretion of metallothionein proteins. Excessive ingestion of Sb by humans can lead to nausea, diarrhea, skin rashes, and respiratory disorders (Figa et al., 2021). The International Agency for Research on Cancer (IARC) reported that Sb trioxide (Sb₂O₃) is a possibly oncogenic substance to humans (IARC, 1989). Further studies by the IARC with Sb₂O₃ revealed unconvincing or unclear results. More recently, the US National Toxicology Program (US-NTP) said that Sb₂O₃ is a human carcinogen based on carcinogenicity tests on model organisms and results from mechanistic investigations (NTP, 2018). Still, the available data related to Sb carcinogenicity on humans are insufficient to evaluate the relationship between people exposed to Sb₂O₃ and human cancer risk.

Remediation of Sb in soil and water ecosystems can be achieved through immobilization, mobilization of Sb using various amendments, and removal through phytoremediation and bioremediation (Diquattro et al., 2021; Hua et al., 2021; Loni et al., 2020; Sun et al., 2017; Zand and Heir, 2020) (Fig. 1). A range of materials has been explored to mitigate Sb risk based on adsorption and immobilization, including activated carbon (Yu et al., 2014), biocomposite materials (Lapo et al., 2019), biochar (Jia et al., 2020), metal oxides (Wang et al., 2019a), mineral sorbents (Zhang et al., 2020), imprinted polymers (Shakerian et al., 2014), and nano zero-valent iron (Huang et al., 2019). Recent literature mentioned the effects of different soil amendments in remediation of Sbimpacted sites in terms of reducing bioavailability and toxicity of the contaminant (Li et al., 2018b; Palansooriya et al., 2020; Rinklebe et al., 2020). Filella and Williams, (2012) and Filella, (2011) previously reviewed, how Sb interact with heterogeneous complexants of the environments which influence their speciation, bioavailability, and further remediation. Furthermore, few recent reviewed discussed the distribution of Sb in various environmental compartments, along with their remediation technologies (Long et al., 2020b; Li et al., 2018b; Artzer et al., 2018). More recently, Nishad and Bhaskarapillai, (2021) reviewed the industrial release of Sb in the environments and their remediation via various physicochemical and biological approaches. However, there

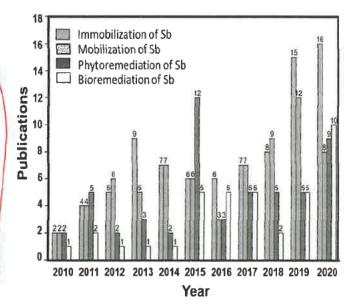


Fig. 1. Number of publications (2010-2020) related to remediation of antimony via various processes.

is no comprehensive review on the biogeochemistry and transformation of Sb related to Sb remediation.

Therefore, the present review integrates the fundamental aspects of Sb transformation and recent developments in understanding Sb speciation and bioavailability related to the remediation of Sb-contaminated terrestrial and aquatic ecosystems. This review presents the current knowledge on the fate of Sb in soils and aquatic environments, contributing to a better cognizance of the environmental hazards of Sb.

2. Origin and sources of Sb contamination

Sb enters the environment via naturally occurring geogenic and anthropogenic pathways. Consequently, it appears in diverse environmental components, including surface and groundwater, soil, sediment, rock, and air (Fig. 2).

2.1. Geogenic sources

Sb is found throughout various environmental compartments (e.g., soil, water, air), and in most biota (Fort et al., 2016; Hiller et al., 2012). Compared to other elements occurring in the earth's crust, the abundance of Sb is low, ranging from 0.2 to 0.3 $\mu g \, g^{-1}$. The abundance of Sb in rocks is usually between 0.2 $\mu g \, g^{-1}$ and several $\mu g \, g^{-1}$, but its higher abundance in shale can reach up to 300 $\mu g \, g^{-1}$ (Zhuang et al., 2018). Sb is present in organic or inorganic form(s) (Qi et al., 2008; Zhuang et al., 2018), predominant in pyrites and several other sulfide minerals (Vikent'eva and Vikentev, 2016). Sb is a scarce element and, in the usual environmental setting, often exists in the forms of stibnite (Sb₂S₃) and Sb bloom (Sb₂O₃) (Zhuang et al., 2018).

Sb concentrations fluctuate under various environmental, geological,

and biogeochemical conditions, due to leaching and its continuous release from ores, minerals, and associated rocks (Ahmad et al., 2014; Okkenhaug et al., 2016). Sb exists in clay-mineral deposits and is associated with the organic matter of coal (Qi et al., 2008). For instance, anomalous Sb in coals is associated with the characteristics of its source areas, including low-temperature magmatic-hydrothermal liquids, volcanic remnants, and groundwater (Qi et al., 2008). Sb in coals is positively correlated with its sulfide form (Qi et al., 2008). Occasionally, pentavalent antimony (Sb(V)) and the more lethal trivalent antimony (Sb(III)) are detected in natural water environments (NHMRG and NRMMG, 2011). Moreover, Sb can be recovered by converting Sb(III) sulfide to its oxide and reducing it with carbon (Coughlin et al., 2020).

Sb is found naturally in sedimentary rocks, soils, and water at 0.15–2 mg kg $^{-1}$, 0.3–8.6 mg kg $^{-1}$, and $<1~\mu g$ mL $^{-1}$, respectively (Pierart et al., 2015). As with most minerals, Sb concentrations are dependent on parent materials, with a typical range of 0.2–10 mg kg $^{-1}$, but usually under 1 mg kg $^{-1}$ (Eschan et al., 2009). Sb levels in clean water and sea water are $<1~\mu g$ L $^{-1}$ and 0.2 μg L $^{-1}$, respectively, and up to 100 μg L $^{-1}$ in the vicinities of an anthropogenically contaminated water (FileBa et al., 2002). Guideline values for acceptable Sb levels in potable water are set at 20 μg L $^{-1}$ (WHO, 2011) and 5.0 μg L $^{-1}$ (Council of the European Enler), 1998). The Australian Drinking Water Guidelines note that concentrations of Sb in potable water should not exceed 3 μg L $^{-1}$ (NHMRC and NRMMC, 2011).

2.2. Anthropogenic sources

The primary source of the hazardous form of Sb (i.e., Sb(V)) mainly originates from industrial activities, such as metal mining and pharmaceutical manufacturing (Zhuang et al., 2018). Other sources include

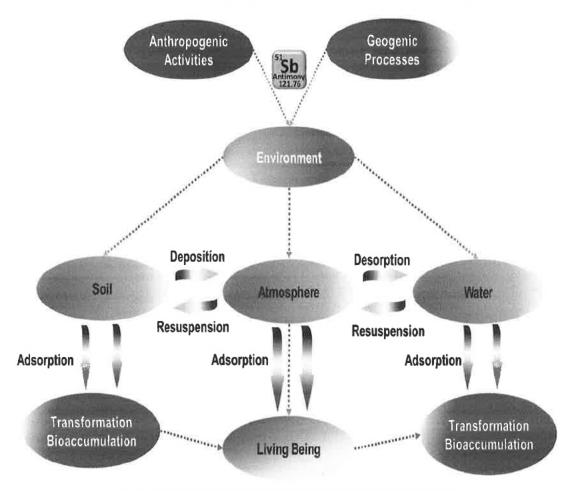


Fig. 2. Antimony sources, transformation, and principal pollution pathways.

sewage sludge (Campos et al., 2019), vehicular emission (Fort et al., 2016), leaching from plastic waste (Chu et al., 2021; Hu et al., 2021), seepage from mining wastes and industrial dumps (Dupont et al., 2016), direct infiltration of leachate from landfill disposal of solid wastes or electronic wastes (Intrakamhaeng et al., 2020), and spillage from industrial metal processing or wood preserving facilities (Herath et al., 2017). Antimonial compounds were first used in ancient times. Sb was found in a 5,000-year-old vase now in the Louvre Museum in Paris (France) and yellow-lead-antimonite-glazed ornamental bricks at Babylon that date from 604 to 561 BCE. Roman Sb-decolourized glass fragments in the Yasmina Necropolis at Carthage provide archaeological evidence of Sb usage prior to the mid-fourth century BC (Schibille et al., 2017). Rom-Sb glasses were recycled and then mixed to produce Rom-Sb-Mn by Romans (Schibille et al., 2017). In modern times, Sb alloys and compounds are also necessary in many manufacturing industries (e. g., semiconductors, motor vehicle batteries, flameproofing materials, pesticides, solder alloys, and fireworks), and in the process of car-tire vulcanization (Bagherifam et al., 2019a; Diquattro et al., 2021; Guo et al., 2018; NHMRC and NRMMC, 2011; Okkenhaug et al., 2016).

Lead bullets used in small-arm shooting ranges are hardened by the addition of 2–8% Sb (Mariussen et al., 2017). As a result, small arm shooting ranges are polluted by the accretion of heavy metal(loid)s into soils, where the key pollutants are lead (Pb), copper (Cu), and Sb. Mariussen et al. (2017) reported that nearly 12 tons of Sb are deposited annually at the Steinsjøen military shooting range in Østre Toten in southern Norway. Sb is mainly present as uncharged Sb(III) (Sb(OH)₃) and anionic Sb(V) (Sb(OH)₆⁻) in soil pore water, which is an easily dissolved oxyanion that depends on soil redox conditions and pH. Mariussen et al. (2017) used iron grit and ferric oxyhydroxide powder, along with limestone, as sorption amendments added to the soil to restrict cation exchange in pore water, which is responsible for Sb(V) mobility. In that study, Sb(V) species dominated in both treated and untreated soil pore-water measurements and the Sb(III) level was less than the limit of detection (0.21 μ g L⁻¹).

The rising environmental plastic and plastic fragments and their management are of global concern (Sridharan et al., 2021a, b; Kumar et al., 2020a; Kumar et al., 2020b). Sb is used as a co-active fire retardant or an additive or catalyst in plastics products (Chu et al., 2021; Turner, 2018; Turner and Filella, 2017). For instance, Turner and Filella (2017) showed that Sb was present in 18% of >800 polymeric, consumable, and daily-use products, including plastics and plastic fragments, rubber, fabrics, fibers, and foams. In 2020, the worldwide production of Sb was assessed to be around 153,000 tons and almost 14% (22,000 tons) of the worldwide production was consumed by the U.S. (United States Geological Survey, 2021). Due to its utilization as an additive in products and its inadequate recycling, the quantity of Sb discharged into natural settings slowly increases (Hu et al., 2021), which poses a threat to health, mainly from the extremely toxic Sb(III) and its associated compounds (He et al., 2019a). Chu et al., (2021) investigated the anthropogenic release of Sb into the environment from polyethylene terephthalate (PET) during different stages of fabrication using a network-model analysis. They reported that the manufacturing of PET released 2,926 tons, followed by its dyeing (2,223 tons) and weaving (908 tons). Disposal of waste PET fibers released 1,108 tons of Sb in landfills. Discarded fiber waste not put in landfills was 872 tons, Mechanical recycling, chemical recycling, and incineration processes released respectively 784, 284, and 25 tons of Sb into the environment (Chu et al., 2021).

Historic Sb mine sites with unmanaged wastes are a serious environmental hazard because they regularly release Sb into the surrounding environment (Dupont et al., 2016; Fu et al., 2016; Hu et al., 2021; Intrakamhaeng et al., 2020) (Table 1). Soils surrounding unrestrained Sb mines are often severely polluted with As and Sb (Wilson et al., 2010). Moreover, precipitation can enable mobilization of soil Sb into surface waters and groundwater (Hiller et al., 2012), which diminishes the quality of the waters and introduces pollutants into the food chain

Table 1
Selected references on different sources and levels of Sb in soil and water

Country	Media	Source of contamination	Concentration level (Soil-mg kg ⁻¹) and (Water-mg L ⁻¹)	Reference
Australia	Soil	Floodplain back- swamp Floodplain levee Control Rhizosphere soil	16.6 ± 1.5 8.3 ± 0.5 0.2 ± 0.1 9.6 - 23.5	Wilson et al. (2014)
Britain	Soil	Former mining and smelting sites	11.94-709.8	Flynn et al. (2003)
China	Soil	Mining	528 ± 259	He et al. (2012)
	Soil	Tailing Dam Forestry	564 ± 252 115 ± 72	
	Soil (roadside	Vegetation	49.75 ± 34.80	
	Soils near	Grassland	39.29 ± 28.36	
	industry)	Coals	2.27	Ql et al. (2008)
		Baosteel Co., Ltd.	3.39 ± 3.59	Yan et al. (2020)
		Gaoqiao Petrochemical Co., Ltd.	2.88 ± 1,39	
		Jiangqiao Domestic Garbage Incineration Plant	2.62 ± 1.85	
	Soils surrounding the tailings	Sb processing plant	1654.462	Zhou et al. (2020)
	Water Samples	Ore concentrate tank	2.168	
		Filter tank	4.415	
		Tailing tank	2.780	
	Soil	Tailing reservoir Mining dump site	3.414 1267.20	Zhou et al. (2019)
	Soil	Mining soil	248-6946,00	Pu et al. (2016)
France	Water Soil	Mining water Mining site	0.0067-0.156 691-1986.00	Elbaz- Poulichet et al. (2020)
Germany	Urban soil	Roadside soil	1.75	Thestorf and Makid, (2021)
italy	Soil	Forest topsoil Abandoned mine	0.54 19-4400	Cidu et al. (2014)
	pore-water		23-1700	
	slag material		0.030	
Corea	runoff Soil	Active firing range	0.8-0.000760 E-6 67.48	Ahmad et al.
witte	UJII.	reave ming range	07.70	(2014)
Macedonia		Former As-Sb mining sites	37.00	Alderton et al. (2014)
Mongolia	Soil Groundwater	Coal Shallow wells	55.20	Qi et al. (2008) Etim, (2017)
Vigeria	Groundwater	(average) Residential	13.5 ± 15.0 to 33.2 ± 36.8 E-6 0-76.0 E-6	Ethn, (2017)
		Commercial Area Industrial	43.0-64.0 E-6 52.0-64.0 E-6	
Norway	Groundwater	Agricultural Small arms shooting ranges	51.0-66.0 E-6 50	Wersin, et al. (2002)
	Runoff	Small arms shooting ranges	5–20	Stromseng et al. (2009)
	Topsoil	Small arms shooting ranges	830	Mariussen et al. (2017)
	Discharge		0.065	
	water Pore-water		19-349	

Table 1 (continued)

Country	Media	Source of contamination	Concentration level (Soil-mg kg ⁻¹) and (Water-mg L ⁻¹)	Reference
Poland	Soil	Historical mining area	0.03-499	Lewinska and Karczewska, (2019)
		Copper Industry	0.12-2.59	
		Shooting Range	10.7-17.0	
		Landfills	0.33-1.55	
	Soil	Former mining sites	12.8-163.00	Lewińska et al (2018)
Slovakia	Soil	Abandoned Sb mines	9861.00	Hiller et al. (2012)
	Water	Abandoned Sb mines	9.30	
Vietnam	Soil	The Mau Due mine	47–95.00	Cappuyus et al. (2021)
Spain	Soil	Mining	14.3-1,090	Murciogo et al (2007)
Scotland	Soil	Upper Spoil	47.77 ± 16.07	Mbadugha et al. (2020)
		Lower Spoil	27.05 ± 5.44	
		Processing	531.47 ± 326.72	
		Surrounding mine	3.66 ± 0.54	
		Burn	3.51 ± 0.99	
		Gully	1.73 ± 0.04	

(Protano and Nannoni, 2018). Mbadugha et al. (2020) noted that this pollution is of concern, when crops are grown on the land. Abad-Valle et al. (2018) reported that pasture plants and agricultural crops accumulate unsafe levels of heavy metals on polluted mine soils.

Mbadugha et al. (2020) investigated As and Sb levels in surface water and soils near a dereliet Sb mine (Louisa mine) in Glendinning, Scotland. They collected 40 soil samples distributed across the study area targeting seven upper spoil heaps, 14 lower spoil heaps, four ore-processing areas, 12 surrounding soils, and three samples located at different depths in the shaft to evaluate the environmental impacts of mining practices on As and Sb levels. The environmental Sb concentration in soils near the Louisa mining site was up to 40 mg kg $^{-1}$, which was higher relative to < 0.5 to 30 mg kg⁻¹ in European soils (Gallagher et al., 1983). High Sb levels were associated with the ore-processing area (i.e., 531.47) \pm 326.72 mg kg $^{-1}$), and found in the region where stibnite-containing ores were processed (Protano and Nannoni, 2018). Levels of Sb diminished with distance from mining and processing zones (i.e., upper spoil had 47.77 \pm 16.07 mg kg⁻¹, lower spoil had 27.05 \pm 5.44 mg kg⁻¹; and locations surrounding the mine had 3.66 \pm 0.54 mg kg $^{-1}$). The water samples from the Gully and Glenshanna Burn sites had 1.73 \pm 0.04 μg ${
m L}^{-1}$ and 3.51 \pm 0.99 ${
m \mu g}$ ${
m L}^{-1}$ dissolved Sb, respectively, much lower than that at a former Sb mine in Scotland (783 µg L-1) (Macgregor et al., 2015); nevertheless, the concentrations were significantly greater than that in unimpacted freshwater ($<1 \mu g L^{-1}$).

Sb production in Australia accounts for nearly two percent of the global share (Red River Resources Ltd, 2020). The Hillgrove mineral field located in the upper Macleay River catchment of northern New South Wales (Australia) has produced over 60,000 tons of stibnite concentrates (i.e., Sb mineral) and gold (Au) (Wilson et al., 2014). Historical mining activity at the Hillgrove mineral field commenced in 1857, ceased in 1921, and recommenced in 1969 (Red River Resources Ltd, 2020). The Hillgrove mineral field was put on care and maintenance in 2016 owing to persistent low Sb prices; it was set to recommence operations by the end of 2020 (Red River Resources Ltd, 2020). Historically poor mine waste management activities have led to the dispersal of Sb-polluted sediments to the coastal floodplain at Kempsey, within 300 km of townships with high population density and intense land use (Ashley et al., 2007). Over 90% of the floodplain exhibits Sb

concentrations above background levels, and the area has above the Australian ecological-investigation values (Tighe et al., 2005). It is a leading anthropogenic source for Sb in Australia (Wilson et al., 2014). Thus, Sb uptake in food crops grown in the region and the resulting human exposure through food-chain transfer is of serious concern.

Sb contamination of groundwater can result from various geogenic sources (i.e., chemical and biological weathering of parent materials, leaching, and wet deposition) or anthropogenic sources (i.e., mining, industrial effluents, and pesticide application) (Etim, 2017). In groundwater, Sb exists mainly in inorganic forms, either in trivalent (Sb (III)) or pentavalent (Sb(V)) species (Etim, 2017). Under varying pH of the media and the available forms of Sb, it may undergo either oxidation or reduction reactions. Soluble forms of Sb tend to travel with water. However, less soluble forms are sorbed onto the surface of clay or soil fragments. Sb can leach from landfills (Intrakamhaeng et al., 2020) and sewage sludge (Campos et al., 2019) into groundwater, surface water, and sediment. Sb leached into sediments can be released into the environment via microbial action and distributed via various processes, as discussed below in detail.

3. Distribution and speciation of Sb in the environment

The occurrence of Sb in different environmental media such as soil, water, and sediments, and its toxicity issues have gained increased attention from both public and the scientific community (Herath et al., 2017; Li et al., 2018b; Zhu et al., 2020). As Sb toxicity depends on its distribution and speciation in such environments, it is crucial to understand these aspects to reduce adverse impacts to ecosystems and human health.

3.1. Distribution in soil

Sb released into soils is naturally retained by aluminum (Al)-, iron (Fe)-, and manganese (Mn)-oxide, while the soluble antimonate exists primarily as Sb(V) in oxidized and alkaline conditions (Ji et al., 2017). Metal(loid) including Sb mobilization in soils depends on multiple factors such as mineral precipitation–dissolution, adsorption–desorption and ion exchange, complexation, pH, Eh, biotransformation transformation, phytostabilization/uptake and leaching (Palansooriya et al., 2020; El-Naggar et al., 2019; El-Naggar et al., 2018). Background and average values of Sb in soils were reported at 0.3–8.6 mg kg $^{-1}$ and 1 mg kg $^{-1}$, respectively, while that in parent rocks was < 1 mg kg $^{-1}$ (Johnson et al., 2005). However, significantly high Sb concentrations in soil (up to 4,400 mg kg $^{-1}$) and soil pore water (up to 1,700 µg L $^{-1}$) samples were reported in an abandoned mine soil (Cidu et al., 2014). The Sb concentration in soils at abandoned mine sites of New South Wales, Australia, was up to 39.4 mg kg $^{-1}$ (Tighe et al., 2005) (Table 2).

The total Sb level in agricultural soils of Spain near an As-Pb-Sb composite (used for lithium-ion battery applications) dumping site was 14.1–324 mg kg⁻¹ (Alvarez-Ayuso et al., 2012), exceeding the highest acceptable limits of Sb in soils recommended by USEPA. Apart from the shooting-range soils analyzed in Norway, several other soils from shooting ranges have been investigated to determine Sb concentration (Table 2) because, as stated, Sb constitutes 2–8% w/w in bullets (He et al., 2019a). In a shooting range in Switzerland, the Sb concentration ranged from 35.0 to 17,500.0 mg kg⁻¹ in the surface soil layer (Johnson et al., 2005). The highest Sb concentration at a military training field in Canada was 570.0 mg kg⁻¹ (Laporte-Saumure et al., 2011) whereas that in the topsoil from a study in Norway was 830.0 mg kg⁻¹ (Mariussen et al., 2017).

3.2. Distribution in the aquatic environment

Sb occurs naturally at low concentrations in unpolluted, natural water bodies. Depending on the physicochemical environmental setting and geographical positioning, the concentration of total dissolved Sb in

Table 2
Distribution of Sb in soil and aquatic environments

Region/ country	Source/site	Species	Concentration (soil-mg kg ⁻¹) and (water- µg L ¹)	Reference
New South, Australia	Mining area Soil	Sb (total)	39.40	Tighe et al. (2005
Xishan, China	Mining area Soil	Sb (total)	5949.20	Qi et al. (2011)
Iran	Agricultural soil	Sb (total)	2.50	Mohammadpour et al. (2016)
China	Mining area soil	Sb (total)	547.00	Conto et al. (2015)
China	Mining area	Sb(III) and (V)	229-1472.00	Wei et al. (2015)
China	Mining area soil	Sb (total)	2675536.00	Nlag et al. (2015
Australia	Wetland soil	Sb (total)	22000.00	Warnken et al. (2017)
Australia	Sb processing industry soil	Sb (total)	7900.00	Ngo et al. (2016)
Italy	Mining area	Sb (total)	4400.00	Chhi et al. (2014
Italy	Mining area	Sb (total)	23-5140.00	Armiento et al. (2017)
Xikuangshan, China	Mining area	Sb (total)	3061.00	l.i et al. (2014)
Australia	Dumping area	Sb (total)	4517.00	Wilson et al. (2013)
Australia	Mining area	Sb (total)	2735.00	Wilson et al. (2013)
Spain	Agricultural soil	Sb (total)	14.1-324.00	Alvarez-Ayuso et al. (2012)
Spain	Mining area soil	Sb (total)	35.00	Pérez-Sirvent et al. (2012)
Spain	Mining area	Sb (total)	5-40.00	Pérez-Sirvent et al. (2011)
Czech Republic	Forest soil	Sb (total)	379.00	Euler et al. (2010
Czech Republic	Agricultural soil	Sb (total)	3.1~131,00	Extler et al. (2010
Switzerland	Shooting field soil	Sb(III) and (V)	21.00	Wan et al. (2013)
Switzerland	Shooting field soil	Sb (total)	21.00	Conesa et al. (2011)
Cologne, Germany	Roadside soil	Sb (total)	6.19	Földi et al. (2018
Xikuangshan, China	Mining area	Sb(III) and (V)	11798.00	Okkenhaug et al. (2011)
Lengshuijiang, China	Dumping area soil	Sb (total)	5949.20	Qi et al. (2011)
Xikuangshan, China	Mining and smelting soil	Sb (total)	5045.00	He, (2007)
Scotland	Mining area	Sb (total)	10-1200.00	Gal et al. (2007)
Spain	Mining area	Sb (total)	225-2450.00	Murciego et al. (2007)
Japan	Smelting industry soil	Sb(V)	2900.00	Taknoka et al. (2005)
Switzerland	Shooting field soil	Sb(V)	35–17500.00	Johnson et al. (2005)
China	Soil	Sb (total)	2.535	Geng et al. (2020
Mau Due mine, North Vietnam	Soil	Sb (total)	31-91.00	Cappuyns and Campen, (2020)
Erme, south west England	Sediment- plastic mixtures	Sb (total)	256-47600.00	James and Tuiner (2020)
Mau Due mine, North	Mining wastes (e.g., slags)	Sb (total)	186-27221.00	Cappuyns and Campen, (2020)
Vietnam taly	Mining area surface water	Sb (total)	0.5-148	Armiento et al. (2017)
	River water	Sb(total	1.2-220.60	Baeza et al. (2010

Table 2 (continued)

Region/ country	Source/site	Species	Concentration (soil-mg kg ⁻¹) and (water- µg L' ¹)	Reference
Yangtze River,		Sb(III)	0.023-0.116,	
China		and (V)	0.047-0.441	
Finland	Groundwater	Sb (total)	0.02-0.82	Lahermo et al. (2002)
Ethiopia	Groundwater	Sb (total)	0.002-1.780	Retmann et al. (2003)
Yunnan Sichuan Province, China	Geothermal waters	Sb (total)	0.23-173.00	Guo et al. (2020)

surface- and ground-water varies from ng L-1 to mg L-1 (Long et al., 2020b). The average Sb concentration is typically $< 1.0 \text{ mg L}^{-1}$ in untapped natural water (Filelia et al., 2002). In recent years, the occurrence of various chemical species of Sb in aquatic systems has been acknowledged (He et al., 2019b; Long et al., 2020b; Li et al., 2018a). Mining activities influence the Sb concentration in soils and natural waters (Long et al., 2020b). The analysis of water samples collected from the Sb-mining site in Xikuangshan (People's Republic of China) revealed 2.0-6,384 µg L⁻¹ oxidized Sb concentrate, considerably higher than the average global Sb concentration (1 µg L⁻¹) (Wang et al., 2011). Of 18 samples taken from the Xikuangshan mining site, the most prevalent Sb species detected was Sb(V), with small Sb(III) concentrations found in four samples (Liu et al., 2010). The drinking water samples collected from the Xikuangshan mine, China, contained 53.6 \pm 46.7 μ g Sb L⁻¹ (Fe et al., 2010). Sharifi et al. (2016) studied Sb content and its flow dynamics in water. The authors proposed that the oxides and hydroxides of ferric iron are the most suitable for Sb reduction via adsorption or coprecipitation which is in agreement with Baeza et al. (2010),

Field trial studies have shown that Sb(III) and Sb(V) contents decrease via dilution, and further their dissolved phase are separate out during downstream movement of the Sb. Resongles et al. (2013) carried out a pH-gradient and oxygen-concentration dependent speciation investigation of suspended Sb particulates in Carnoules mine water (southern France). The results showed that 70-100% of Sb particulates belonged to Sb(III), and < 30% Sb belonged to Sb(V). Future studies need to consider the effects of unpredictable weather conditions on the distribution of Sb in contaminated sites, particularly from mine sites. For illustration, intense rainfalls alter Sb distribution in environmental compartments, by increasing the Sb level in oceans via rivers and sediment transport off agricultural fields (Armiento et al., 2017). Alderton et al. (2014) reported that surface-water samples contained more Sb (average 0.6–2.1 μg L⁻¹) than groundwater samples. Similarly, variation was reported among surface and bottom layers of fresh waters. Wu et al. (2011b) compared the key species of soluble Sb in the estuary of the Yangtze River with a nearby freshwater site. The findings showed that Sb(V) and Sb(III) levels in surface waters ranged from 0.121 to 0.567 μg L⁻¹ and 0.029-0.736 μg L⁻¹, respectively. The bottom layer had the lowest concentrations of Sb(V) (0.047-0.441 µg L-1) and Sb(III) (0.023-0.116 μg L-1). Comparable results have been documented by others (Asaoka et al., 2012; Jablońska-Czapla et al., 2015).

3.3. Chemical forms and speciation

The Sb concentration in the environment is dependent on the soil redox potential (Eh), pH, the presence and content of Fe- and Mn-containing ores or minerals, soil cation exchange capacity, humic acids (HAs), and dissolved organic matter (DOM) (Herath et al., 2017; Rinklebe et al., 2020). In soil and sediment, oxide and hydroxides of Fe and Mn play key roles in the adsorption of Sb, particularly Sb(III). For example, Fe and Mn accelerate the oxidation rate of Sb(III) to Sb(V) and change the concentration of both the species (Xu et al., 2011). However,

soluble Sb compounds usually persists in four distinct chemical states [Sb(-III)/Sb(0)/Sb(III)/Sb(V)] (Moreno-Andrade et al., 2020; Li et al., 2018b). The chemical states of Sb determine its transformation, mobilization, and immobilization. Sb(III) has a lower solubility and mobility than Sb(V) (Johnson et al., 2005; Wu et al., 2011b). Although Sb(III) has lower water solubility than Sb(V), the former has a stronger affinity towards Fe and Mn hydroxides, and HAs in terrestrial systems (Liu et al., 2015). Generally, compared to Sb(III), higher Sb(V) concentrations occur in natural aquatic environments (Wilson et al., 2010; Filella et al., 2002; Fu et al., 2016). Aquatic Sb concentrations are influenced by Sb sorption mechanisms, Sb mobilization or immobilization, pH, Eh, cooccurring metals ions, and DOM (Frohne et al., 2011; Herath et al., 2017; Rinklebe et al., 2020). Natural organic matter may also regulate aqueous phase Sb concentrations (Filella and May 2005; Filella and Williams. 2012). Buschmann and Sigg, (2004) showed that Sb(III) was bound via carboxylic and phenolic groups. >30% of overall Sb(III) can be bound with aquatic HAs, and the process is strongly influenced by low ionic strengths of the media (Nakamaru and Peinado, 2017).

Furthermore, the pH of a specific medium has a critical role to play in Sb speciation in that medium. In the pH range of 1.0–11.0, Sb(III) generally occurs in its hydroxide form such as Sb(OH)₃, SbO(OH), and HSbO₂, whereas under extreme, low pH (<1.0), Sb(III) persists in the cationic form SbO⁺/Sb(OH)₂⁺ (Bergmann and Koparal, 2007). At higher pH (>11.0), the predominant Sb(III) forms exist as anions SbO₂⁻ or Sb (OH)₄. In the pH range of 1.0–14.0, Sb(OH)₆⁻ is the dominant form of Sb(V), while at the lowest pH (<1), Sb(V) primary exists in the SbO₂⁺ form (Long et al., 2020b). Sb(V) was found to be predominant and exists in the form of Sb(OH)₆⁻ in the drainage water samples collected from the Xikuangshan Sb mine, China (Liu et al., 2010).

Computational modeling predicted that, in anaerobic water at pH ranges from 1.4 to 11.8, the Sb(III) form exists as the hydroxy complex (Sb(OH)3). However, under aerobic, aquatic conditions, the negatively charged (Sb(OH)6") is the predominant species of Sb(V) when pH > 2.7 (Filella and May 2003). Nevertheless, in anaerobic and aerobic waters, both forms [Sb(III), Sb(V)] exist (Filella et al., 2002), and this is ascribed to microbial activities, chemical-facilitated changes in Sb(III) - Sb(V) in anaerobic waters, or mixing with concurring waters (Kulp et al., 2014; Li et al., 2013). Sb(III) oxidation, facilitated by Fe and Mn, is controlled by several ores or minerals and trace metals present in natural environments (Lenz et al., 2006; Multani et al., 2016). Sb(III) oxidation using hydrogen peroxide (H2O2) was studied under measured environmental concentrations of Pb²⁺, Zn²⁺, Cu²⁺, and Mn²⁺ (Elleonet et al., 2005). The results revealed that the presence of trace elements enhanced the reaction kinetics over a wide-ranging pH. Some microorganisms are able to transform Sb(III) and Sb(V) forms into several organic and inorganic compounds via diverse mechanisms, such as fluxion in concentrations of Sb(III), oxidation and reduction of Sb(III) and Sb(V), respectively, and methylation of Sb(III) (Filella et al., 2007; Li et al., 2016). Lehr et al. (2007) showed Sb(III) oxidation using wild and mutant varieties of Agrobacterium tumefaciens. In anaerobic water sediments, Kulp et al. (2014) showed bacteria-mediated Sb(V) reduction via a respiratory route using lactate and acetate as energy sources. However, microbeassisted Sb oxidation is in its infancy and needs more research to understand the mechanisms involved (Li et al., 2013).

4. Biogeochemical processes of Sb in the environment

The global biogeochemical cycle for Sb is poorly documented in comparison to other elements in the literature. Some studies have attempted to describe the atmospheric emissions of Sb (Fort et al., 2016; Ijjima et al., 2008; Tian et al., 2011; Tian et al., 2014); however, the transformation pathways and quantities of Sb among different environmental compartments (e.g., soil, aquatic bodies, biota, and atmosphere) have not been well defined. Additional information about the dispersal and speciation of Sb in the natural environment is necessary in assisting scientists to fill this gap. Meanwhile, an up-to-date

understanding of different biogeochemical processes of Sb in soil and aquatic environments is essential, which is described in this section (Fig. 3).

4.1. Biogeochemical processes of Sb in the soil

Soil characteristics highly influence the biogeochemical processes of Sb (Constantino et al., 2018; Frohne et al., 2011; Natasha et al., 2019; Rinklebe et al., 2020), which in turn determine Sb speciation, retention versus mobility in soil, plant uptake, and transfer to the water environment (Herath et al., 2017). Such biogeochemical processes include adsorption-desorption, dissolution, leaching, (co)precipitation, redox transformations, and interactions of Sb with microorganisms (Fan et al., 2020; Natasha et al., 2019; Zhu et al., 2020).

The adsorption process is a major retention mechanism for Sb in soil. Sb is adsorbed by different soil constituents, including clay minerals and other inorganic adsorbents and organic adsorbents. For instance, clay minerals, such as montmorillonite, have a high adsorption capacity for Sb (Kabata-Pendias, 2011). The weathering of clay minerals and breaking of their edges stimulate the adsorption of Sb onto clay mineral surfaces (Herath et al., 2017; Natasha et al., 2019).

Inorganic adsorbents, such as Fe-, Mn-, and Al-(hydr)oxides (also constituents of soil clays e.g., layered-aluminosilicate clay minerals) affect Sb adsorption in soil. Minerals like hematite, ferrihydrite, and goethite contain Fe-oxides, which participate in immobilizing Sb in soil. As a result, several studies have attempted to use Fe-based adsorbents to remediate of soils contaminated with Sb. For instance, goethite adsorbed Sb(III) over a pH range from 3 to 12, and Sb(V) at pH < 7 (Filella and Williams, 2012). Zero-valent, Fe-functionalized-carbon nanotubes is a competent adsorbent material for Sb(III) and Sb(V) (Mishra et al., 2016). In particular, the authors showed high adsorption of Sb(III) (250 mg g-1), which was attributed to the high surface area of nano-zero-valent Fe particles. Naturally occurring Mn oxides (birnessite) and Al oxides (gibbsite) and their hydroxides, are common minerals in soils. Mn and Al oxihydroxides can interact with Sb(III) and Sb(V), and reduce Sb mobility in agricultural soils. Belzile et al. (2001) described the oxidation mechanism of Sb(III) by Mn oxides under anaerobic conditions as follows:

$$MnO2(S) + Sb(OH)3(aq) + H2O(t) \rightarrow Mn(OH)2(aq) + H3SbO4(aq)$$
(1)

MnO₂ is more capable of oxidizing Sb(III) than Fe (hydr)oxides (Belzile et al., 2001; He et al., 2019b). A combination of Fe-Mn oxide materials showed greater adsorption capacity for Sb(III) than Fe (hydr) oxides alone (1.76 mmol g⁻¹ vs. 0.83 mmol g⁻¹). Sb(III) oxidation to Sb (V) by Mn oxides, which facilitated Sb(V) adsorption by Fe (hydr)oxides, was the underlying mechanism (Xu et al., 2011). Direct adsorption of Sb (V) by Mn (hydr)oxides, however, is not yet well documented. Sb(V) adsorption on surfaces of gibbsite was described (Eq. II and III) by He et al. (2019b):

$$Al - OH_{(S)} + H_{6(aq)}^+ \leftrightarrow Al - (OH)_{2(S)}^+$$
 (2)

$$2AI - OH_{(S)} + Sb(OH)_{6(uq)}^{-} \rightarrow (AIO)_2Sb(OH)_{4(S)}^{-} + 2H_2O_{(1)}$$
 (3)

The surface charge of Al oxides will become prevalently positive in acidic environments (Kosmulski, 2009), which stimulates electrostatic forces that interact with anionic forms of Sb(V) (He et al., 2019b; Xu et al., 2001). Regarding Sb(III) adsorption, (hydr)oxides of Fe and Al have a lower adsorption capacity than Mn (hydr)oxides. However, due to the higher abundance of (hydr)oxides of Fe and Al in soil than Mn (hydr)oxides, the former usually govern the behavior of Sb(III) (Wilson et al., 2010).

Sb adsorption is also influenced by soil organic matter (SOM) (Inam et al., 2019). Soil HAs accommodated a considerable amount of Sb(III) (up to 34% of total soil Sb level) (Buschmann and Sigg, 2004; Ettler et al., 2007; Tscrenpil and Liu, 2011), demonstrating a high affinity

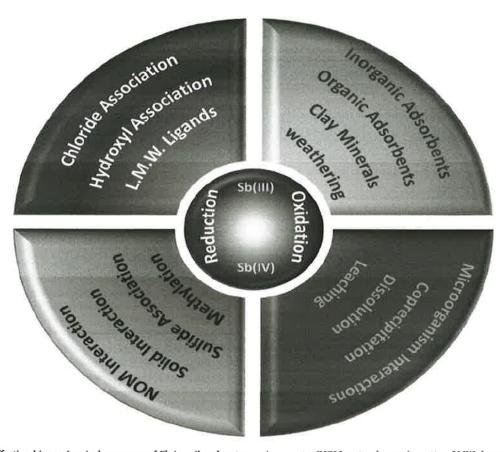


Fig. 3. Factors affecting biogeochemical processes of Sb in soil and water environments. (NOM: natural organic matter; LMW: low molecular weight).

between Sb(III) and SOM. The postulated complexation mechanisms of Sb(III) with functional groups of OM were described by Wilson et al. (2010), including the generation of anionic Sb complexes with carboxy groups and ligand substitution with the Sb-center. HAs can also stimulate Sb(III) oxidation to Sb(V). Humic substances were reported to adsorb more Sb(III) than Sb(V) (Inam et al., 2019); however, the underlying mechanism explaining this disparity is not yet well documented, and further investigations are required in this aspect (Wilson et al., 2010).

Other biogeochemical processes of Sb in soil include the dissolution and leaching of Sb from Sb-bearing primary minerals (e.g., Sb2O3 and Sb₂S₃) and co-existence of Sb(III) and Sb(V) forms in soil (Cappuyns et al., 2021; Herath et al., 2017; Shan et al., 2020). A recent study in Vietnam found that wastes at the Mau Due Sb mine, where the primary Sb mineral was stibnite (Sb₂S₃), leached up to 1.7% of its total Sb content (191-15,699 mg kg-1) to water, and the Sb release increased at a pH range 7.5-12.8 (Cappuyns et al., 2021). Several factors were reported to accelerate the dissolution and leaching processes of Sb, including photo-irradiation, low-molecular-weight-dissolved organic acids, total SOM content, sulfur oxidation, soil pH, and soil Eh (Frohne et al., 2011; Fu et al., 2016; He et al., 2019b; Rinklebe et al., 2020; Shan et al., 2020). Dissolution of Sb is always associated with dynamic alteration in soil biogeochemical conditions, enhancing its mobility in soil and facilitating Sb translocation from the soil profile into water bodies. For instance, soil under flooded, anaerobic conditions commonly has a high dissolution rate of Fe oxides, accompanied by the release of DOM (Rinklebe et al., 2016; Shaheen et al., 2014). Consequently, flooding conditions can induce dissolution of Sb-Fe oxides minerals and Ca-antimonate (Rinklebe et al., 2020; Zhu et al., 2020). Additionally, the dissolution of dissolved organic carbon under flooded condition can result in increased Sb release owing to the strong Sb-organic molecule association. However, soil flooding can also accelerate the adsorption of Sb due to Sb(V) reduction to Sb(III), which has higher affinity to SOM (Grob et al., 2018). These contrasting results are mainly attributed to the variable conditions of soil pH, site-abundant soil minerals including Fe and Al oxides, and the induced changes of soil Eh, levels of soil-dissolved organic carbon, and sulfur chemistry.

(Co)precipitation of Sb may also occur in soil via binding of Sb with compounds of amorphous hydroxides and Ca (Randelović et al., 2020; Wilson et al., 2010). Sequential dissolution and re-precipitation of Sb (OH)3 reportedly controlled Sb availability in soil surrounding a mine site in Vietnam (Cappuyns et al., 2021). However, (co)precipitation of Sb can only have a major role when soil Sb concentration is elevated to a level that allows the (co)precipitation process. It is supposed to have a marginal role in soils with low total Sb concentration. In soils with low total Sb concentration, adsorption-desorption processes are the major mechanisms for immobilization of Sb (Herath et al., 2017). The biogeochemical processes of Sb also can be affected by interactions between soil microorganisms and Sb. For instance, arbuscular mycorrhizal fungi, bacteria, and archaea can induce changes in Sb speciation and bioavailability in soils, thereby affecting plant uptake of Sb (Girolkar et al., 2021; Wei et al., 2016). Similarly, Fe- and S-oxidizing and reducing bacterial species are likely to impact Sb transformations in soil. The interactions between Sb and soil microorganisms are discussed in detail in Section 5.

4.2. Biogeochemical processes of Sb in aquatic environments

Biogeochemistry of Sb in aquatic systems has been rarely studied, and this field lacks sufficient experimental data (Fu et al., 2016). In aqueous solution, Sb can form complexes with low-molecular-weight ligands and interact with colloidal and solid phases of inorganic or organic substances; these processes can be affected by living organisms (Filella and Williams, 2012; Sun et al., 2017). Sb(III) and Sb(V) can readily hydrolyze in aquatic systems, particularly under alkaline conditions, while Sb species are more stable in acidic solutions (Filella and

Williams, 2012). Interactions of Sb with low-molecular-weight ligands, such as hydroxyl, chloride, sulfides, and organic substances, vary widely according to the pH of aqueous solutions. For instance, SbO₂⁺ dominates under acidic conditions, but, when the solution pH turns to be neutral or alkaline, Sb can interact with hydroxyls to form Sb(OH)₆⁻ as the predominant species (Filelia et al., 2002; He et al., 2019b; Herath et al., 2017). In terms of Sb interactions with chloride, limited data are available. It is not yet evident whether chloride or hydroxy-chloride containing forms of Sb(III) and Sb(V) are formed in seawater.

Under an oxic environment, Sb(V) is the abundant form in solution, while Sb(III) is the common form under anoxic conditions. Sb can interact with sulfur under anoxic conditions, forming both soluble (e.g., SbS₂²⁻) and/or insoluble (e.g., Sb₂S_{3(s)}) species, which is dependent on pH of the system (Filella et al., 2002; Polack et al., 2009). Interactions of Sb with organic ligands having O-bearing functional groups, such as alcohols, phenols, and carboxylic acids, also have an important role in Sb mobility in aquatic environments (Dousova et al., 2015). He et al. (2019b) reported that up to 85% of total Sb in lake water was complexed with organic substances, such as fulvic and HAs. In aqueous solutions, Sb (III) can be associated with carboxylic and hydroxy carboxylic acid functional groups within a pH range between 3 and 9 (Tella and Pokrovski, 2009). However, the binding of Sb(V) by organic ligands is not well documented.

Understanding the biogeochemical processes of Sb in water would allow the application of suitable amendments to reduce the toxicity of Sb to living organisms. Manganese oxides can be efficient in the immobilization of Sb(III) in water environments under anoxic conditions via adsorption and oxidation mechanisms (Fig. et al., 2019b). However, compared to other toxic elements, Sb has received much less attention in research that elucidates detailed biogeochemical processes and mechanisms and water remediation techniques.

In conclusion, various factors affect the biogeochemical processes of Sb in soil and water, as discussed previously in this review and represented in (Fig. 3). Therefore, future research is warranted to ascertain Sb interactions with solid phases and natural OM, speciation of methylated Sb, and induced biogeochemical changes in Sb species under different oxic-anoxic conditions in soil and water environments.

5. Bioavailability and toxicity of Sb to biota

Due to its toxicological nature, Sb contamination and translocation through the ecosystem can pose severe threats to biota and human health. To assess the potential risks, regulatory guidelines based on total metalloid concentration in soils are commonly used (Greenberg et al., 2014). However, binding with Fe and Mn oxides, adsorption, precipitation, and complexation in soil solution may hinder the mobility of Sb and other metalloids. Thus, the assessment based on total metal(loid) concentration including Sb may overestimate of the potential risks (Bagherifam et al., 2019b; Bolan et al., 2008). Therefore, considering the dynamic process of bioavailability is important.

5.1. Bioavailability of Sb

To be bioavailable, Sb must be in its mobile form and should be readily available for biological uptake to pose a risk on target organs (Kuppusamy et al., 2017). Water-soluble Sb can be readily available for plant uptake, and excessive accumulation can lead to toxic effects (Corrales et al., 2014; Tschan et al., 2010). The bioavailability of Sb in the soil environment is mainly governed by several factors as represented in (Fig. 4). A plot scale study by Zhong et al. (2020) revealed that Sb is more easily adsorbed by plant roots in alkaline soil (pH 8.39) than acidic soil (pH 4.91). Moreover, the bioavailability of Sb increases when soil Eh decreases (Zhu et al., 2020). Management practices also affect Sb bioavailability. For instance, prolonged flooded conditions, which are required for paddy cultivation, cause favorable conditions for excessive Sb uptake by plants (Zhu et al., 2020).

Nakamaru and Peinado (2017) reported that Sb availability to plants could increase by 2-4 times by increasing SOM content. Sb bioavailability is also influenced by the presence of ions (PO₄³⁻, NO₃⁻, CO₃²⁻, and SO₄²) via competitive adsorption and co-precipitation (Zhu et al., 2020). Associated dissolution of Fe and Mn (hydr)oxides can also increase Sb bioavailability. Wan et al. (2013) reported decreased available Sb levels initially with flooded conditions due to increased proportion of Sb(III) under reduced conditions, but subsequently increased owing to the dissolution of Fe and Mn oxides. This resulted in a 10-fold increment in shoot Sb level of Lolium perenne L. (perennial ryegrass) but an 80% decrease in Holcus lanatus L. The results indicate the importance of the influence of oxidation state and species on Sb uptake. The presence of NO₃ in soil may inhibit the reductive dissolution of Fe minerals, thus lowering the bioavailability of Sb in paddy soils (Zhang et al., 2021b). Soil microorganisms, such as bacteria, archaea, and fungi, can mediate the bioavailability of Sb by inducing speciation changes (Li et al., 2016; Long et al., 2020a). For example, inoculation of plant roots with fungal species of arbuscular mycorrhizas could promote Sb uptake (Pierart et al., 2015; Wei et al., 2016).

Most plants have a higher affinity for absorbing Sb(III) than Sb(V) (Huang et al., 2012; Ji et al., 2017; Ren et al., 2014). Several methods are deployed for determining Sb availability in soil, such as Sb measurement in pore water, one-step- or successive-extraction methods (Bagherifam et al., 2019b), and the thin-film-diffusion-gradient technique (DGT) (Luo et al., 2010). Compared to total Sb in soil, low levels of water-soluble Sb were found in Sb-contaminated soil in Germany (0.5%) (Lintschinger et al., 1997), the United Kingdom (<1%) (Flynn et al., 2003), and Jiangxí Province, People's Republic of China (0.52–3.51%) (Liu et al., 2007), which confirm its low bioavailability.

An in vivo test by Denys et al. (2012) showed that the relative bioavailability of Sb varied from 2 to 6% in mining- and smeltingcontaminated soil. In the same study, no Sb was detected in biological endpoints of kidney, liver, and bones of humans from the smeltercontaminated soils, implying low bioavailability in terrestrial ecosystems. Several methods are commonly used to determine in vitro amounts of bio-accessible Sb concentrations. These methods include the Simplified Bioaccessibility Extraction Test (SBET), the Physiologically Based Extraction Test (PBET), and Unified BARGE Method (UBM) (BARGE is the BioAccessibility Research Group of Europe) (Denys et al., 2012; Li et al., 2014). Sb concentrations were reported ranging from 19 to 60,000 mg kg⁻¹ in soil samples from a contaminated mining site at Glendinning, Scotland, UK, with the bio-accessible amount in gastric extraction ranging from 5.05 to 9.11%, as determined by the UBM method (Denys et al., 2012). An average Sb level of 3,061 mg kg⁻¹ was reported in soils from the Xikuangshan Sb-mining area, China, with average bio accessibility values of 5.89 \pm 6.44%, 7.83 \pm 9.82%, 3.03 \pm 3.53% determined by SBET, PBET (gastric), and PBET (intestinal) extraction, respectively (Li et al., 2014). Wilson et al. (2014) reported Sb levels of 0.2-16.6 mg kg-1 in the Macleay River Floodplain in northern New South Wales, Australia, with bioaccessibility < 3%, as measured by SBET. Using the UBM test, an investigation on a football field contaminated with mine tailings in France demonstrated Sb bio accessibility of less than detection limits, although the total Sb concentrations ranged from 157 to 363 mg kg⁻¹ (Pascaud et al., 2014). Lower bio accessibility of Sb in the intestinal phase (1-28%) than the gastric phase (6-83%) was shown in soils collected from a shooting range in Australia (Sanderson et al., 2012). In most cases, low bio accessibility of Sb in soils can be attributed to entrapment of Sb by Fe oxyhydroxides, the presence of sulfides in soils, or the association of Sb with the residual fraction of soil (Herath et al., 2017). High Sb bioavailability can cause toxicities in animals, plants and humans (He et al., 2019a).

5.2. Toxicity to plants and microorganisms

Sb existing in soil solution can be readily taken up by plants, potentially resulting in adverse impacts to the plants (Maresca et al.,

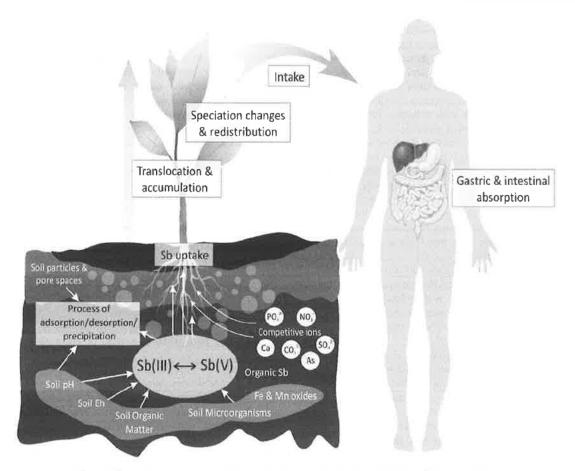


Fig. 4. Schematic representation of the major factors affecting bioavailability and uptake of Sb.

2020). The Sb-associated, plant-toxic effects are growth retardation, repression of photosynthesis, uptake and assimilation of substances that are generally not taken up, and reduced synthesis of some metabolites and hormones (Back et al., 2014; Feng et al., 2013; Zhou et al., 2018). Zhou et al. (2018) studied Acorus calamus (common name: calamus), a tall perennial herb, and observed that both Sb(III) and Sb(V) species adversely affected the synthesis of leaf photosynthetic pigments (i.e., chlorophyll a and b and carotenoids), plant height, dry weight, and net photosynthetic rate. Previously, Feng et al. (2016) revealed that the various species of Sb would be alter the morphology of plant roots and adversely impacted development of the root system. Feng et al. (2020) reported that Sb(III) increased the biosynthesis of abscisic acid in rice plants, which can impede cell division, elongation, and growth. Furthermore, various species of Sb persuaded oxidative stress, nevertheless rice crop accelerated the actions of ascorbate peroxidase (APX) and superoxide dismutase (SOD) to counter the oxidative stress. Plant hormone synthesis can also be altered as a result of Sb(III) uptake, affecting root morphology (Zhu et al., 2020). Furthermore, various species of Sb persuaded oxidative stress, nevertheless rice crop accelerated the actions of ascorbate peroxidase (APX) and superoxide dismutase (SOD) to counter the oxidative stress.

Sb toxicities on microorganisms have been well studied and reported. Kataoka et al. (2018) reported a strong selective pressure on As (III)-oxidizing bacterial populations in a soil bacterial community, induced by various Sb species. The same study revealed that the coexistence of Sb with As has an important role on As redox reactions in the environment. Although these toxic effects are present, some microorganisms are capable of remediating Sb in impacted environments (Waog et al., 2011). Various chemical processes of microorganisms enable them to survive in Sb-rich environments (Li et al., 2021), which can be identified as efflux, methylation, Sb(III) oxidation, and Sb(V)

reduction (Li et al., 2016). For example, in yeast, the accretion of the lethal, intracellular form of Sb(III) was reduced by preventing its entry into cells or, if it entered cells, its active expulsion. Also, a harmless form could be made inside the cell space (Wysocki et al., 2001). Further, microbial-assisted oxidation of Sb(III) transmuted Sb(III) to its oxidized form (Sb(V)), which is less toxic and more immobilized, thereby assisting bioremediation of Sb-contaminated soil and allowing for its safe disposal (Li et al., 2019).

5.3. Risk to animals and humans

Sb is documented as a reckoned oncogenic hazardous contaminant. It displays multifarious compound specific geno and cyto-toxicity and there is adequate proof to suppose Sb containing compounds to be a potential human carcinogen. Exposure to Sb or Sb-derived complexes; through dermal, oral, and inhalation, by animals and humans can have grave adverse health impacts (Bagherifam et al., 2019b; Wu et al., 2013a). Sb compounds have an affinity toward thiol groups (i.e., sulfhydryls) of glutathione and proteins. They retained for an extended duration and disturb enzymatic activity, and adversely affecting several organ systems, including lungs, heart, liver, and kidneys (Li et al., 2016; Wang et al., 2018). An early study showed that inhalation of Sb(III) oxide leads to lung cancer in female mice (Groth et al., 1986). Back et al. (2014) studied the response of Asian earthworm (Perionyx excavates) exposed to elevated Sb levels. Negative effects on the earthworm's survival and morphological abnormalities (i.e., surface-casting activity) were reported (Back et al., 2014).

After exposure to Sb, systemic, immunological, neurological, reproductive, genotoxic, and developmental effects and cancer, have been well documented by the United States Public Health Service (US-PHS) (US-PHS, 1992). Sudden-infant-death syndrome (SIDS) is found to be

related to the inhalation of toxic gases (e.g., stibines) that originate from methylation of Sb species by the fungus Scopulariopsis brevicaulis in polyurethane-foamed crib mattresses (Richardson, 1994). However, studies are also available indicating no causal link to SIDS with Sb biovolatilization (Cullen et al., 2000; Jenkins et al., 2000). A health risk analysis found that the hazard quotient (HQ) values of Sb in vegetables ranged from 1.61 to 3.33 (Zeng et al., 2015). A HQ is defined by dividing the chronic daily intake by the mean reference dose; if HQ value is higher than 1, it appears that there are obvious health risks. In this study, the chronic daily intake was above the safe limit prescribed by the Food and Agriculture Organization (FAO) and WHO. These results indicated that long-term utilization of vegetables from Sb contaminated soils might jeopardize the health of inhabitants near the Xikuangshan mine in Hunan, China (Zeng et al., 2015). However, there are few data existing to justify the adverse impact of Sb contamination and their level of toxicity to human beings as there is always a co-exposure of As as well to the working staffs exposed to Sb during mining and smelting activities (Nishad and Bhaskarapillai, 2021; NTP, 2018). Most of the investigations and research lack adequate size of the sample and appropriate control population, and that make the outcomes inconsistence. However, there are few recent animal investigations and reports revealed the toxicity of Sb to living organisms (Schildreth et al., 2020; Saerens et al., 2019; Agency for Toxic Substances and Disease Registry, Atlanta, 2019).

6. Risk management of Sb in contaminated environments

Both bioavailable and immobilized fractions of Sb are found in the soil matrix. Numerous studies have considered the bioavailable fraction of Sb in soil and found it important (Majzlán et al., 2016; Nakamaru and Peinado, 2017). Therefore, actions need to be taken to manage the bioavailable fraction of Sb in the soil, mainly where agricultural practices are undertaken. However, most experiments that have focused on risk management of Sb in contaminated soils have been restricted to laboratory research, with limited field applications due to several factors (Fig. 5).

6.1. Immobilization using soil amendments

The application of materials (e.g., clay minerals, biochar, organic composts, and fly ash) to a metal(loid) including Sb contaminated soil can reduce flow and transport of Sb in the soil and its availability for plants, owing to the strong sorption of such materials (Bolan et al., 2021; Kumar et al., 2020c; Palansooriya et al., 2020). Among various amendments, chemical additives provide the best immobilizing potential compared to others, including biochar (Kumar et al., 2021a; Doherty et al., 2017; Silvani et al., 2019). In some cases, biochar increased Sb mobilization in soil; therefore, the use of biochar needs further consideration before its application in Sb contaminated soils (Flua et al., 2021; Rinklebe et al., 2020).

Several reports have demonstrated the effective immobilization of Sb by ferrous sulfate, which induces Sb(V) reduction to Sb(III), and Sb(III)

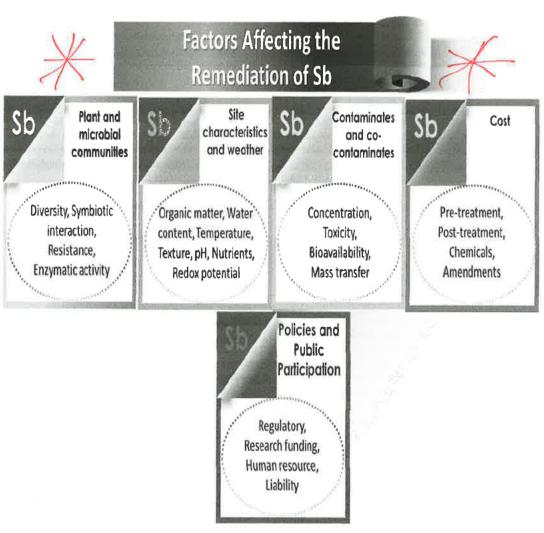


Fig. 5. Factors affecting field-scale remediation of Sb contaminated sites.

became firmly bound with iron oxide/hydroxide over a wide-ranging pH (Almäs et al., 2019; Tandy et al., 2017). Biochar modified with Fe-Mn immobilized Sb in a contaminated soil and thereby, reducing Sb phytoavailability in the soil (Wang et al., 2019c). Moreover, the amendment of Fe-modified biochar to agricultural soil increased Sb retention and soil fertility (Wang et al., 2019c). The influence of urea on Sb bioavailability in a paddy-field soil was investigated by Zhang et al. (2021b). The results demonstrated that, at the beginning (15-45 days after treatment), Sb bioavailability extracted by ammonium sulfate increased compared to the control soil (9.5-9.8 vs. 10.1-14 mg kg⁻¹) due to the higher soil pH releasing adsorbed and organic-bound Sb from the soil. After 120 days of experimentation, reduced Sb bioavailability was observed due to the enhanced affinity of Sb toward iron oxides present in the soil. Immobilization of Sb, as shown in various studies, and its efficiencies, are given in Table 3(a). Although the application of such soil amendments results in the high retention of metal(loids), some drawbacks have not been rectified. In addition, any change in soil conditions (e.g., pH) can activate the bioavailability of metal(loids) (Wuana and Okieimen, 2011). Most studies have focused only on one aspect (i.e., Sb immobilization). However, it is important to investigate the associated changes in activities of soil enzymes and fertility.

6.2. Mobilization using soil leaching

The mobilization of Sb in contaminated soils has not been carried out to a great extent, mainly due to the high cost of the chelators. Various chelating agents, both organic and inorganic, such as ethylenediaminetetraacetic acid (EDTA), polyacrylic acid, acetic acid, oxalic acid, citrate, ethylenediamine tetra(methylene phosphonic acid), ammonium oxalate, ammonium acetate, calcium chloride, ammonium nitrate, and disodium hydrogen phosphate, have been assessed (Filella, 2011). A low level of Sb was leached from the soil using gentle extractants, and a better-leached yield of Sb was achieved by organic extractants, except EDTA (Filelia, 2011). Tan et al. (2018) subjected lightly, moderately, and highly contaminated soils to a leaching process with different extractants; the results showed that the efficiency of extractants in leaching Sb from the soils decreased in the order of citric acid, tartaric acid, EDTA, hydrochloric acid, disodium hydrogen phosphate, and calcium chloride. Interestingly, in a highly Sb contaminated soil (6,876 mg kg⁻¹), water leached up to of 94.8 mg kg⁻¹ Sb (Tan et al., 2018). Water and disodium hydrogen phosphate mobilized almost equal amounts of Sb (Soil No. 1: 0.6 mg L-1 and Soil No. 2: 0.3 mg L-1) (Cappuyns and Van Campen, 2020). The efficiencies of mobilization data are presented in Table 3(b).

6.3. Phytoremediation

Phytoremediation is a lengthy but cost-effective, and aesthetically accepted approach used to remediate environmentally harmful metal (loids)/pollutants including Sb (Antoniadis et al., 2021; Fuke et al., 2021; Kumar et al., 2021c; Mensah et al., 2021; Prabha et al., 2021). It is a greener and environmentally sustainable remediation technique, than other physicochemical methods for metal mobilization and immobilization (Gunarathne et al., 2020). Most data are from laboratory (pot) experiments, in which the soil has been spiked with Sb. However, spiking does not represent the same mineralogical composition of the Sb-derivatives and leads to rapid bioavailability. Müller et al. (2013) conducted a 7-week pot experiment with Sb. A clean soil spiked with 5, 10, and 16 mg kg $^{-1}$ of Sb and an Sb-contaminated soil (14 mg kg $^{-1}$) from an abandoned mine area were used. The authors also studied the effect of a co-contaminant (arsenic) added at a rate of 20 mg kg⁻¹ on the studied plant species (Pteris vittate, a fern). A significant amount (232.6 mg kg⁻¹) of Sb was taken up by the plant when grown in the pots spiked with 5 mg kg⁻¹ and grown with the co-contaminant, but only 52.9 mg kg⁻¹ was taken up when the plants were exposed only to Sb. Sb accumulated in the roots (230 mg kg⁻¹) rather than in the shoots (2.6 mg kg⁻¹). Sb in the plants grown in the contaminated mining soil was 20.1 mg kg⁻¹ (Müller et al., 2013). In contrast, for *Sorghum bicolor* (sorghum), the Sb concentration in shoots was about 2-fold higher than that in roots (translocation factor was 2.11) (Zand and Heir, 2020). The translocation factor (TF) is calculated by dividing the Sb concentration in shoots by that in roots. If TF > 1, the plant can be used as a good phytoremediator (Antoniadis et al., 2021). Qi et al. (2011) compiled data from 31 plants belonging to 21 families to determine their phytoremediation potential; they reported Sb concentrations in the plants ranging from 3.92 to 143.69 mg kg⁻¹. The TF values of some plants are displayed in Table 3 (c). A few of the plants, such as *Barbarea verna* (a type of cress), *Sorghum bicolor* (sorghum), and *Nicotiana* (tobacco) (Family: Solanaceae) had TF > 1, and plants might be potential phytoremediators for Sb.

6.4. Microbial remediation

Microbial communities in soils are crucial biotic factors in the remediation of potential toxic elements such as Sb, via bio-oxidation and bio-reduction, which change the toxicity, mobility, and labile nature of Sb by altering its chemical properties (He et al., 2019b; Jevasundar et al., 2021). Numerous Sb-oxidizing bacteria species have been reported. For instance, Acinetobacter sp., Stenotrophomonas sp., Comamonas sp., Shinella sp., Hydrogenophaga sp., Variovorax sp., Variovorax sp., and Flavihumibacter stibioxidans; these strains facilitate Sb (III) oxidation to Sb (V) under aerobic conditions (Han et al., 2016; He et al., 2019b). One strain of Shinella sp. (NLS1) and one strain of Ensifer sp. (NLS4), isolated and screened from Sb-polluted soil, showed optimum Sb (III) oxidation in agar media with and without the presence of yeast extract (Choi et al., 2017) (Table 3(d)). However, some strains belonging to the order Bacillales (Lai et al., 2016; Lai et al., 2018) transmuted Sb(V) to Sb (III) under anoxic conditions. A new strain belonging to the Sinorhizobium genus was isolated from Sb-contaminated sediment; it reduced Sb (V) to Sb (III) in the presence of acetate under anaerobic conditions (Nguyen and Lee, 2014). Another bioremediation process in the microbial community is the methylation of inorganic Sb. Methylated Sb exhibits less toxicity than pentavalent Sb and trivalent Sb (He et al., 2019b). The production of monomethyl Sb, dimethyl Sb, and trimethyl Sb by microbial communities has been well documented (Andrewes et al., 2000; Hartmann et al., 2003). However, no studies have been performed to demonstrate the immobilization of microbially methylated Sb in soil using amendments.

6.5. Integrated risk management

An integrated approach incorporates a set of remediation strategies deployed to remediate contaminated environments (Kumar et al., 2021b; Sun et al., 2020). Several researchers (e.g., Chirakkara et al., 2015; Girolkar et al., 2021; Sánchez et al., 2020) have studied this topic. Couto et al. (2015) used phytoremediation, coupled with electrokinetics and a phosphorus amendment to remove Sb from the soil using Indian mustard (Brassica juncea) and ryegrass. The addition of a phosphorus amendment increased Sb uptake by ryegrass by 25% and by Indian mustard by 30% than the plants alone. The phosphorus amendment enhanced Sb desorption from the soil due to the ionic repulsion between phosphate ions and Sb, and Sb bioavailability increased with a low-intensity electric field (Couto et al., 2015). Effects of TiO2 nanoparticles on the phytoremediation of Sb using Sorghum bicolor was undertaken by Zand and Heir (2020). The plant was exposed to varying dosages of TiO2 nanoparticles (from 0 to 1000 mg kg-1). Increased total plant biomass, plant uptake and translocation of Sb in the pots containing 50-250 mg kg⁻¹ TiO₂ relative to those in the control soil (without TiO2) were reported (Zand and Heir, 2020).

7. Summary and future research needs

Environmental Sb contamination is triggered by natural processes

Table 3
The remediation process of antimony (Sb (III) and Sb (V)).

Environmental matrix	Initial concentration	Amendment	Efficiency	References
(a) Immobilization Retention efficienc				
Soil/\$lag (50.00 g/50.00 g)	72.7.0 mg kg ⁻¹	FeSO ₄ (4.00 g)	99.25%	Almäs et al.
Soil/Slag (45,00 g/10.00 g)	72.70 mg kg ⁻¹	Peat soil (45.00 g)	39.15%	(2019)
Soil	79.24 mg kg ⁻¹	Fe-Mn biochar (0.5% w/w)	33.80%	Wang et al.
		Fe-Mn biochar (1.00% w/w)	40.50%	(2019c)
		Fe-Mn biochar (2.00% w/w)	43.50%	Conne
Low TOC soil	100.00 mg kg ⁻¹	Pristine biochar (20.00% w/w)	12.00%	Silvani et al.
LOW TOC SOIL	100.00 mg kg			
W.1 man . II	010.00 - 11	Fe-enriched biochar (20.00% w/w)	47.00%	(2019)
High TOC soil	210.00 mg kg ⁻¹	Pristine biochar (20.00% w/w)v	90.00%	
		Fe-enriched biochar (20.00% w/w)	60.00%	
Soil (4.00 g)	445.165 $\mu g \ kg^{-1}$ leached	Zero valent Fe ⁰ (1:19 ratio to soil)	6.66%	Tandy et ol.
	concentration of Sb in unamended	FeSO ₄ (1:19 ratio to soil)	98.32%	(2017)
		Viro-Soil 1 (1:4 ratio to soil)	76.52%, 47.62%	
		Fe ⁰ : Viro-Soil 1: soil (4:19:76)	79.37%	
		Fe ⁰ : Viro-Soil 2: soil (4:19:76)	36.24%	
		FeSO ₄ : Viro-Soil 1: soil (4:19:76)	97.19%	
		FeSO ₄ : Viro-Soil 2: soil (4:19:76)	96.74%	
		reso ₄ . viio-soii 2. soii (4.19.70)		
			Decrease in Sb leaching	WY C 5 54
Fly ash	$2.04 \pm 0.06 \text{ mg L}^{-1}$	Laboratory iron-rich residuals (LIR-1	3.53–6.53% and 4.68–10.58% by	Wang et al.
		and LIR-2) (1.75% w/w)	LIR-1 and LIR-2 respectively	(2019b)
Soil (70.00 g)	Column leached concentration of	Kaolinite 8,00% (w/w)	46.50%	Doberty et al.
	Sb in unamended: 35.34 µg	Kaolinite 10.00% (w/w)	44.70%	(2017)
		Zero valent Fe 1.00% (w/w)	67.00%	
		Zero valent Fe 3.00% (w/w)	82.30%	
		Zero valent Fe 5.00% (w/w)	72.30%	
			82.00%	
		Ferrihydrite 1.00% (w/w)		
		Ferrihydrite 3.00% (w/w)	92.60%	
		Ferrihydrite 5.00% (w/w)	93.30%	
		Ferric chloride 1.00% (w/w)	74.90%	
		Ferric chloride 3.00% (w/w)	54.60%	
(b) Mobilization/leaching process			Leaching percentage/amount	
Soil (5.00 g)	1234.70 mg kg ⁻¹ *	EDTA (25.00 mL, 0.05 M) and few drops	Mean: 7.464% (n = 12)	Zhang et al.
Soil (2.50 g)	1234.70 mg kg ⁻¹ *	of 1.00 M NaOH	Mean: 1.741% (n = 12)	(2018)
Soil (2.50 g)	1234.70 mg kg ⁻¹ *	Na ₂ HPO ₄ (25.00 mL, 0.10 M)	Mean: 0.721% (n == 12)	A CONTROL OF A
3011 (2.30 g)	1204.70 116 18	Ultra-pure H ₂ O (25.00 mL)	Wedi: 0.72170 (II - 12)	
0 11 4 .	0.01.40.00 . 1.11-1			eres to the control of the control o
Soil 1 and 2 (1.00 g)	Soil 1: 47.00 mg kg ⁻¹	Demineralized H ₂ O (10.00 mL)	Soil 1: 0.612 mg L ¹	Cappuyns et a
	Soil 2: 95.00 mg kg ⁻¹		Soil 2: 0.284 mg L ⁻¹	(3031)
Soil 1 and 2 (1.00 g)	Soil 1: 47.00 mg kg ⁻¹	Na ₂ HPO ₄ (10.00 mL, 0.10 M)	Soil 1: 0.635 mg L ⁻¹	
	Soil 2: 95.00 mg kg ⁻¹		Soil 2: 0.326 mg L ¹	
Lightly contaminated soil (1.00 g)	145.00 mg kg~ 1	CaCl ₂ (20.00 mL, 0.05 M)	0.87 mg kg ⁻¹	Tan et al.
		Na ₂ HPO ₄ (20.00 mL, 0.50 M)	7.39 mg kg ⁻¹	(2018)
		HCl (20.00 mL, 0.50 M)	7.10 mg kg ⁻¹	
		EDTA (20.00 mL, 0.10 M)	8.68 mg kg ⁻¹	
		Tartaric acid (20.00 mL, 0.04 M)	21.70 mg kg ⁻¹	
		Citric acid (20.00 mL, 0.10 M)	34.40 mg kg ⁻¹	
Moderately contaminated soil (1.00 g)	720.00 mg kg ⁻¹	CaCl ₂ (20.00 mL, 0.05 M)	5.32 mg kg ⁻¹	
		Na ₂ HPO ₄ (20.00 mL, 0.50 M)	22.20 mg kg ⁻¹	
		HCl (20.00 mL, 0.50 M)	17.70 mg kg ⁻¹	
		EDTA (20.00 ml, 0.10 M)	57.10 mg kg ⁻¹	
		Tartaric acid (20.00 mL, 0.04 M)	64.90 mg kg ⁻¹	
		Citric acid (20.00 mL, 0.10 M)	117.00 mg kg ⁻¹	
lighly contaminated soil (1.00 g)	6876.00 mg kg ⁻¹	CaCl ₂ (20.00 mL, 0.05 M)	59.40 mg kg ⁻¹	
riigiliy containnated son (1.00 g)	0870.00 IIIg Kg			
		Na ₂ HPO ₄ (20.00 mL, 0.50 M)	210.00 mg kg ⁻¹	
		HCl (20.00 mL, 0.50 M)	478.00 mg kg ⁻¹	
		EDTA (20.00 mL, 0.10 M)	1217.00 mg kg ⁻¹	
		Tartaric acid (20.00 mL, 0.04 M)	902.00 mg kg ⁻¹	
.00		Citric acid (20.00 mL, 0.10 M)	1449.00 mg kg ⁻¹	
Mineral soil (100.00 g)	72.70 mg kg ⁻¹	Deionized H ₂ O (20-30.00 mL)	57.68 mg L ⁻¹	Almās et al.
-		-		(2019)
(c) Phytoremediation		Plant species	Concentration or	
			Translocation factor (TF)	
Mining area soil	5949.20 mg kg ⁻¹	Hippochcaete ramosissima	Plant: 98.23 mg kg ⁻¹	Qi et al. (201)
AUTHUR STES 2011	3743.20 IIIg Kg			Su er ur fron
		Rubia leiocaulis	Plant: 66.15 mg kg ⁻¹	
		Pteris henryi	Plant: 45.53 mg kg ⁻¹	
		Pteris vittate	Plant: 49.16 mg kg ⁻¹	
		Debregeasia edulis	Plant: 46.15 mg kg-1	
Mining area soil	2904.00 and 44.40 mg kg ⁻¹ (aqua	Agrostis capillaris (Plant collected at	Root: 402.00 μg g ⁻¹	flech of id-
	regia and EDTA extraction,	mine area)	Shoot: 69.00 µg g ⁻¹	(2012)
	-	samely the ball	,	CALLERY.
	respectively	Assessed a sealth to Contract to the first	TF: 0.172	
		Agrostis capillaris (Plant potted in a mine	Root: 98.75 μg g ⁻¹	
		soil (20%) and quartz sand (80%)	Shoot: 115.68 μg g ⁻¹	
		mixture)	TF: 1.17	

(continued on next page)

Table 3 (continued)

nvironmental matrix	Initial concentration	Amendment	Efficiency	References
		Agrostis capillaris (Plant potted in a mine soil (50%) and quartz sand (50%)	Root: 190 μg g ⁻¹ Shoot: 45 μg g ⁻¹	
		mixture)	TF: 0.24	
ining area soil	152.00 and 19.20 mg kg ⁻¹ (aqua	Роа аппиа	Root: 19.00 µg g ⁻¹	Bech et al.
	regia and EDTA extraction,		Shoot: 4.60 μg g ⁻¹	(2012)
	respectively)		TF: 0.24	
		Veronica persica	Root: 5.70 µg g ⁻¹	
			Shoot: 1.70 µg g ⁻¹	
			TF: 0.30	
		Sonchus asper	Root: 3.60 μg g ⁻¹	
			Shoot: 2.60 µg g ⁻¹	
			TF: 0.72	
		Barbarea verna	Root: 1.90 µg g ⁻¹	
			Shoot: 3.10 μg g ⁻¹	
			TF: 1.63	
ining area soil	55	Bubblejaceae Buddleja	Shoot: 14.24 mg kg ⁻¹	Levresse et a
				(2012)
		Ruscaeae Dasylinin	Shoot: 26.59 mg kg ⁻¹	
		Asteraceae Gymnosperna	Root: 20.42 mg kg ⁻¹	
			Shoot: 22.52 mg kg^{-1}	
			TF: 1.10	
		Pinaceae Pinus	Shoot: 26.63 mg kg ⁻¹	
oil from waste tailing area	=	Solanaceae Nicotiana	Root: 4.72 mg kg ⁻¹	Levresse et ni
			Shoot: 191.32 mg kg ⁻¹	(2012)
			TF: 40.53	
		Cupressaceae Junipesus	Shoot: 250.64 mg kg ⁻¹	
		Asphodelaceae Asphodelus	Root: 447.52 mg kg ⁻¹	
		•	Shoot: 29.42 mg kg ⁻¹	
			TF: 0.07	
		Myrthaceae Eucalyptus	Shoot: 71.57 mg kg ⁻¹	
oil from abandoned mine	14.00 mg kg ⁻¹	Pteris vittata	Root: 25.00 mg kg ⁻¹	Müller et al-
			Shoot: 0.10 mg kg ⁻¹	(2013)
			TF: 0.004	,,
uartz sand spiked Sb	5.00 mg kg ⁻¹	Pteris vittata	Root: 49.00 mg kg ⁻¹	
			Shoot: 3.90 mg kg ⁻¹	
			TF: 0.08	
uartz sand spiked Sb	10.00 mg kg ⁻¹	Pteris vittata	Root; 68.00 mg kg ⁻¹	
	20100 118 118	A DOI OF PIECES OF	Shoot: 3.40 mg kg ¹	
			TF: 0.05	
uartz sand spiked Sb	16.00 mg kg ⁻¹	Pteris vittata	Root: 63.00 mg kg ⁻¹	
and an addition of the control of th	10.00 mg ng	T to a fitting	Shoot: 9.60 mg kg ⁻¹	
			TF: 0.15	
uartz sand spiked Sb and 5.00 mg kg^{-1}	5.00 mg kg ⁻¹	Sorghum bicolor	Root; 84.00 mg kg ⁻¹	
As (co-contaminant)	3.00 mg kg	Sorgram bacotor	Shoot: 4.40 mg kg ⁻¹	
76 (Co-Contaminant)				
uartz sand spiked Sb and 10.00 mg	5.00 mg kg ⁻¹	Caraban binder	TF: 0.05	
kg ⁻¹ As (co-contaminant)	3.00 mg kg	Sorghum bicolor	Root; 94.00 mg kg ⁻¹	
kg As (co-containmant)			Shoot: 4.10 mg kg ⁻¹	
on 20 00 hard the disable and 20 on	F 00 11	0-1-1:-1-	TF: 0.04	
uartz sand spiked Sb and 20.00 mg	5.00 mg kg^{-1}	Sorghum bicolor	Root: 230.00 mg kg ⁻¹	
kg ⁻¹ As (co-contaminant)			Shoot: 2.60 mg kg ⁻¹	
II	5.00 11		TF: 0.01	
oil contaminated with Sb	5.00 mg kg ⁻¹	Sorghum bicolor	Root: 31.78 mg kg ⁻¹	Zand and Hei
			Shoot: 67.06 mg kg ⁻¹	(2020)
			TF: 2.11	
		0		
mining soil	-	Setaria viridis	Rhizospheric soils: 311 mg kg ⁻¹	Long et al.
mining soil	-	Setaria viridis	Leaves: 7.58 mg kg ⁻¹	Long et al. (2018)
mining soil	-	Setaria viridis	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹	
mining soil	-	Setaria viridis Leucospermum gracile	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹	
mining soil			Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹	
mining soil			Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹	
mining soil			Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹	
mining soil		Leucospermum gracile	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹	
mining soil		Leucospermum gracile	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹	
mining soil		Leucospermum gracile	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹	
mining soil		Leucospermum gracile Cornus canadensis	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soits: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soits: 50.4 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹	
mining soil		Leucospermum gracile Cornus canadensis	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹ Rhizospheric soils: 452 mg kg ⁻¹	
mining soil		Leucospermum gracile Cornus canadensis	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹ Rhizospheric soils: 452 mg kg ⁻¹ Leaves: 174 mg kg ⁻¹ Shoots: 222 mg kg ⁻¹	
mining soil		Leucospermum gracile Cornus canadensis Alloteuthis subulatus	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹ Rhizospheric soils: 452 mg kg ⁻¹ Leaves: 174 mg kg ⁻¹ Shoots: 222 mg kg ⁻¹ Rhizospheric soils: 132 mg kg ⁻¹	The second secon
mining soil		Leucospermum gracile Cornus canadensis Alloteuthis subulatus	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹ Rhizospheric soils: 452 mg kg ⁻¹ Leaves: 174 mg kg ⁻¹ Shoots: 222 mg kg ⁻¹ Rhizospheric soils: 132 mg kg ⁻¹ Leaves: 22.3 mg kg ⁻¹	The second secon
mining soil		Leucospermum gracile Cornus canadensis Alloteuthis subulatus A. thunb	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹ Rhizospheric soils: 452 mg kg ⁻¹ Leaves: 174 mg kg ⁻¹ Shoots: 222 mg kg ⁻¹ Rhizospheric soils: 132 mg kg ⁻¹ Leaves: 22.3 mg kg ⁻¹ Shoots: 37.5 mg kg ⁻¹	
mining soil		Leucospermum gracile Cornus canadensis Alloteuthis subulatus	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹ Rhizospheric soils: 452 mg kg ⁻¹ Leaves: 174 mg kg ⁻¹ Shoots: 222 mg kg ⁻¹ Shoots: 222 mg kg ⁻¹ Chaves: 22.3 mg kg ⁻¹ Shoots: 37.5 mg kg ⁻¹ Rhizospheric soils: 132 mg kg ⁻¹ Rhizospheric soils: 215 mg kg ⁻¹	The second secon
mining soil		Leucospermum gracile Cornus canadensis Alloteuthis subulatus A. thunb	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹ Rhizospheric soils: 452 mg kg ⁻¹ Leaves: 174 mg kg ⁻¹ Shoots: 222 mg kg ⁻¹ Rhizospheric soils: 132 mg kg ⁻¹ Leaves: 22.3 mg kg ⁻¹ Shoots: 37.5 mg kg ⁻¹ Rhizospheric soils: 215 mg kg ⁻¹ Leaves: 70.3 mg kg ⁻¹	The second secon
mining soil		Leucospermum gracile Cornus canadensis Alloteuthis subulatus A. thunb S. bunge	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹ Rhizospheric soils: 452 mg kg ⁻¹ Leaves: 174 mg kg ⁻¹ Shoots: 222 mg kg ⁻¹ Rhizospheric soils: 132 mg kg ⁻¹ Leaves: 22.3 mg kg ⁻¹ Rhizospheric soils: 135 mg kg ⁻¹ Leaves: 25.3 mg kg ⁻¹ Leaves: 25.3 mg kg ⁻¹ Shoots: 37.5 mg kg ⁻¹ Rhizospheric soils: 215 mg kg ⁻¹ Leaves: 70.3 mg kg ⁻¹ Shoots: 31.8 mg kg ⁻¹	
mining soil		Leucospermum gracile Cornus canadensis Alloteuthis subulatus A. thunb	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹ Rhizospheric soils: 50.4 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹ Rhizospheric soils: 452 mg kg ⁻¹ Leaves: 174 mg kg ⁻¹ Shoots: 222 mg kg ⁻¹ Rhizospheric soils: 132 mg kg ⁻¹ Leaves: 22.3 mg kg ⁻¹ Shoots: 37.5 mg kg ⁻¹ Shoots: 37.5 mg kg ⁻¹ Leaves: 70.3 mg kg ⁻¹ Leaves: 70.3 mg kg ⁻¹ Shoots: 31.8 mg kg ⁻¹ Rhizospheric soils: 105 mg kg ⁻¹ Rhizospheric soils: 105 mg kg ⁻¹	
mining soil		Leucospermum gracile Cornus canadensis Alloteuthis subulatus A. thunb S. bunge	Leaves: 7.58 mg kg ⁻¹ Shoots: 4.40 mg kg ⁻¹ Rhizospheric soils: 74.8 mg kg ⁻¹ Leaves: 38.4 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Shoots: 24.2 mg kg ⁻¹ Leaves: 23.3 mg kg ⁻¹ Shoots: 61.5 mg kg ⁻¹ Rhizospheric soils: 452 mg kg ⁻¹ Leaves: 174 mg kg ⁻¹ Shoots: 222 mg kg ⁻¹ Rhizospheric soils: 132 mg kg ⁻¹ Leaves: 22.3 mg kg ⁻¹ Rhizospheric soils: 135 mg kg ⁻¹ Leaves: 25.3 mg kg ⁻¹ Leaves: 25.3 mg kg ⁻¹ Shoots: 37.5 mg kg ⁻¹ Rhizospheric soils: 215 mg kg ⁻¹ Leaves: 70.3 mg kg ⁻¹ Shoots: 31.8 mg kg ⁻¹	

Table 3 (continued)

Environmental matrix	Initial concentration	Amendment	Efficiency	References
			Rhizospheric soils: 316 mg kg ⁻¹	
			Leaves: 55 mg kg ⁻¹	
			Shoots: 43.3 mg kg ⁻¹	
		Eleusine indica	Rhizospheric soils: 53.8 mg kg ⁻¹	
			Leaves: 37.7 mg kg ⁻¹	
			Shoots: 39.2 mg kg ⁻¹	
		Miscanthus sinensis	Rhizospheric soils: 1561 mg kg ⁻¹	
			Leaves: 60.2 mg kg ⁻¹	
			Shoots: 66.7 mg kg ⁻¹	
		A. lavandulaefolia	Rhizospheric soils: 87.1 mg kg ⁻¹	
			Leaves: 28.8 mg kg ⁻¹	
			Shoots: 68.6 mg kg ⁻¹	
(d) Microbial remediation				
Isolation site		Microbial species	Transformation efficiency/	
The stanian musified from Ch	E00.00 -34	Chinelle on CNI (21) and Providence	quantity	QUARTE STOCKING W.
The strains purified from Sb contaminated soil	500.00 µМ	Shinella sp. (NLS1) and Ensifer sp. (NLS4)	Within 1.5 days 500.00 µM of Sb (III) oxidized to Sb (V)	Choi et al. (2017)
contaminated son	3069.30 mg kg ⁻¹	Sinorhizobium sp.	>94.00% of Sb (V) reduced to Sb	(2017)
	3009.30 mg kg	Зикинавокий вр.	(III)	
Strain isolated from Sb mining area soil	Site 1: 1240.00 mg kg-1	Bacillus sp. S3	100.00% of 100.00 µM Sb (III)	Nguyen and
at 4 sites	Site 2: 1550.00 mg kg ⁻¹	,	oxidized to Sb (V) in 2 days	Lee, (2014)
	Site 3: 650.00 mg kg ⁻¹		ondition to be (v) in a days	200, 220, 17,
	Site 4: 90.00 mg kg ⁻¹			
Strain isolated from Sb mining area soil	Site 1: 1240.00 mg kg ⁻¹	Cupriavidus sp. S1	100.00% of 50.00 μM Sb (III)	Li et al. (2018a
at 4 sites			oxidized to Sb (V) in 12 days	
	Site 2: 1550.00 mg kg ⁻¹	Moraxella sp. S2	50.00% of 50.00 μM Sb (III)	
	Site 3: 650.00 mg kg ⁻¹		oxidized to Sb(V) in 14 days	
	Site 4: 90.00 mg kg ⁻¹			
Microbial consortia enriched with Sb	· =	Microbial consortia	Sb(V) (1 mM) was completely	Park and Lee,
contaminated soil			reduced within 1 day	(2019)
Bacterial strains isolated from the Sb	14000 mg kg ⁻¹	Arthrobacter sp.	Sb(V) oxidized to Sb ₂ O ₃ or SbO ₄	Nam et al.
contaminated soils by selective		Aminobacter sp.		(2018)
enrichment	1 -M Cham	Minchiel annualis (Dans)	30004 - Cal-(m)	201
Indigenous microbial community	1 mM Sb(HI)	Microbial consortia (Paracoccus,	~100% of Sb(III) was oxidized in	Zhang et al.
		Rhizobium, Achromobacter and Hydrogenophaga)	36 h	(2021a)

^{*} Values calculated, TOC: Total Organic Carbon, Viro-Soil: Red mud derivative.

and anthropological actions. Elevated levels of Sb in the environment pose serious risks to ecosystems and human health in terms of potential toxicity and carcinogenicity. The existence and dispersal of elevated Sb levels in different environmental compartments are of particular concern in many countries. Although Sb exists in various oxidative-state compounds in both organic and inorganic forms, it occurs principally as inorganic Sb(III) and Sb(V) species in terrestrial and aquatic ecosystems. Sb(III) compounds are more mobile and bioavailable, and thus, more toxic to human and ecosystem health, than Sb(V) species. Chemical speciation of Sb plays a significant role in determining its biogeochemical behavior involving dissolution, sorption/desorption, bioavailability, toxicity, and accumulation in organisms. Various techniques have been used to remove Sb from the environment or reduce its toxicity, such as Sb immobilization using amendments (e.g., activated carbon, metal oxides, inorganic minerals), mobilization via soil washing and phytoremediation, and microbial remediation (e.g., Sb(III) methylation). As each technique has its limitations, an integrated approach is often used for sustainable remediation outcomes.

The present review identified some key knowledge gaps regarding Sb contamination and risk management, which warrant the following future research:

- I. More data regarding Sb distribution and speciation in different environmental media should be collected to establish the global biogeochemical cycle of Sb.
- II. There is relatively little information on Sb levels for different chemical species in environmental media, biota (including flora and fauna), dietary sources, and the human system. The threshold at which Sb has an effect needs to be determined. Therefore, more attention should be paid to assess the ecotoxicology of Sb to fully protect ecosystems and human health.

- III. Chemical and microbiological interactions of Sb often have been studied independently in different environmental settings under controlled conditions. However, under realistic environmental conditions, the combined effects of the two factors are much more complicated and need more research.
- IV. As bioremediation and phytoremediation are increasingly used as green and sustainable techniques for Sb remediation, Sb biosensors needs to be developed to predict remediation outcomes. Future studies regarding biosensors (e.g., Sb(III)-oxidizing bacteria) would shed more light on the remediation efficiency.
- V. Field studies with an integrated approach for the remediation of Sb-contaminated terrestrial and aquatic ecosystems also need to be considered.

Declaration of Competing Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

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N. Bolan et al. Environment International 158 (2022) 106908

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